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Review Article

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Review

Potential Synergistic Supplementation of NAD+ Promoting Compounds as a Strategy for Increasing Healthspan

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Abstract: Disrupted biological function, manifesting through the hallmarks of aging, poses one of the largest threats to healthspan and risk of disease development, such as metabolic disorders, cardiovascular ailments, and neurodegeneration. In recent years, numerous geroprotectors, senolytics, and other nutraceuticals have emerged as potential disruptors of aging and may be viable interventions in the immediate state of human longevity science. In this review, we focus on the decrease in nicotinamide adenine dinucleotide (NAD+) with age and the supplementation of NAD+ precursors, such as nicotinamide mononucleotide (NMN) or nicotinamide riboside (NR), in combination with other geroprotective compounds, to restore NAD+ levels present in youth. Furthermore, these geroprotectors may enhance the efficacy of NMN supplementation while concurrently providing their own numerous health benefits. By analyzing the prevention of NAD+ degradation through the inhibition of CD38 or supporting protective downstream agents of SIRT1, we provide a potential framework of the CD38/NAD+/SIRT1 axis through which geroprotectors may enhance the efficacy of NAD+ precursor supplementation and reduce the risk of age-related diseases, thereby potentiating healthspan in humans.

Keywords: aging; longevity; supplements; geroprotectors; SIRT1; NAD+; NMN; resveratrol; nutraceuticals; age-related diseases; flavonoids; senolytics; healthspan

1. Introduction

The doubling of life expectancy in the last century can be attributed to progressive leaps in health and clinical care in the human population. However, at the expense of longevity comes the increased risk of disease and other physical ailments. Being the main risk factor for the ever-increasing incidence of neurodegeneration, cardiovascular disease, cancer, and other medical ailments, aging has come to the forefront as a new clinical target within itself, aiming to serve as a preventative measure for developing diseases [1]. Aging can be characterized as universal, intrinsic, progressive, and deleterious to the health of an individual. A multimodal onslaught of the body's physiological processes may be the culprit, whether it be through environmental damage, a decline in endogenous protective mechanisms, or a pure breakdown of the physiological process due to wear and tear [2]. Numerous pathways are also implicated and may break down or become dysfunctional due to aging, resulting in neurodegenerative disorders, as an example [3]. The societal burden of these diseases and age-related pathologies is ever-compounding and must be addressed with novel interventions to improve quality of life for the aging population. The conundrum of aging and the associated risk of developing age-related diseases has the

propensity to affect every individual and is further affected and potentially accelerated by other biological and environmental risk factors.

Targeting the hallmarks of aging exists as a model which would allow for interventions ameliorating these processes: genomic instability, telomere attrition, epigenetic alterations, loss of proteostasis, deregulated nutrient sensing, mitochondrial dysfunction, cellular senescence, stem cell exhaustion, and altered intercellular communication. Each intervention is thought to focus on these hallmarks or even have a multimodal crossover effect ameliorating the biological processes of aging [4]. Appropriate, widespread, and easily executable interventions are key factors in combating the newly defined epidemic of aging and age-related disorders. In the field of orthomolecular medicine, the use of supplementation as a therapeutic approach to slow the process of aging is highly applicable when selecting and formulating geroprotective treatments. Longevity supplements have shown exciting potential for improving healthspan, and research into longevity supplements is one of the most rapidly advancing fields in aging science today. However, further human studies are still warranted [5]. Human trials utilizing regenerative or senolytic compounds to target skin aging, in particular, have shown improvement in the biological response to environmental stressors, such as UV and oxidative damage, which often accelerate the process of aging [6–8]. Age-related deficits in cognition also arise as viable targets of longevity boosting and geroprotective supplementation. These deficits can be attributed to excessive neuroinflammation, oxidative damage, and metabolic disruption. Nutraceutical interventions, particularly those targeting these numerous disruptive pathways, have shown promise in ameliorating age-related cognitive decline [9]. Administration of combinations of micronutrients and other antioxidant compounds has also shown promising results in promoting longevity and slowing accelerated aging in mice [10]. One compound in particular, nicotinamide adenine dinucleotide (NAD+), is relevant in many biological processes as a cofactor for crucial metabolic functions. These sensitive pathways are interrelated with sirtuins, a family of signaling proteins involved in metabolic regulation, and PARP1, an enzyme that detects DNA damage and facilitates repair pathways. These may be disrupted by external factors and/or intrinsic breakdown due to aging, leading to an increased risk for many disorders, cognitive decline associated with aging, and neurodegenerative disorders such as Alzheimer's and Parkinson's disease [11]. Multiple cohort studies spanning various age groups have shown significant decreases in NAD+ levels with age. This dysregulation identifies NAD+ as a potential target for age-related pathologies [12]. In model organisms, NAD+ precursor administration has resulted in delayed muscle atrophy, improved neurodegenerative pathologies [13], and restored metabolic function [14]. The clinically relevant evidence regarding the targeting of NAD+ to replenish endogenous stores and invigorate the NAD+ salvage pathway is compounding, raising the question of how NAD+ administration may be able to affect aging or age-related deficits in humans. Given the importance of maintaining NAD+ levels for optimal health and longevity, restoring NAD+ levels is a promising and feasible approach in promoting health during aging.

Here, we review NAD+ metabolism and its molecular interactions with other longevity-promoting pathways. Numerous studies have analyzed the effects of oral NAD+ precursor administration on specific disease states, muscle performance, metabolic processes, and other metrics. However, the longevity-promoting effect of NAD+ precursor administration can be further optimized and enhanced by the supplementation of NAD+ enhancing and supporting compounds [15]. The longevity-boosting potential of repurposed drugs, such as rapamycin and metformin, has been discussed. However, as data from clinical trials are either lacking or only very sparsely available, they cannot be recommended for clinical routine. Ingredients show promise in the context of longevity, but require further research on their synergistic effects on the NAD+/SIRT1/CD38 axis [16]. With no proven pharmaceutic options available, we will discuss other potent ingredients and nutraceuticals that may influence and modulate the NAD+ axis involving SIRT1 and CD38 in biological mechanisms, to offer further geroprotection by combating the aging process and, additionally, providing each of the nutraceutical's own health benefits.

2. NAD+ Homeostasis

Nicotinamide adenine dinucleotide (NAD+) is one of the primary coenzymes in metabolic processes and is involved with numerous other pathways, such as energy expenditure, metabolic and stress adaptations, and circadian rhythm maintenance. NAD+ levels sharply decline with age, and this decline can be attributed to the activity of CD38, an enzyme responsible for the degradation of NAD+, which disrupts the NAD+ synthesis pathways during the course of aging [17]. NAD+ homeostasis in the body is critical for optimal biological function. NAD+ consuming enzymes function to convert NAD+ to nicotinamide (NAM) and occupy a specific role in biological aging pathology. They are implicated as interventional targets for geroprotection, such as the enzyme CD38, the sirtuins (SIRT) deacetylases, poly [ADP-ribose] polymerase 1 (PARP1: involved in the DNA damage response (DDR)), and the neuronal degenerating and NAD+ draining factor SARM1 [18,19]. NAD+ can be synthesized de novo from nicotinic acid (NA), nicotinamide riboside (NR), and nicotinamide mononucleotide (NMN) or can be salvaged through the NAD+ salvage pathway, which is crucial for recycling the metabolites of biochemical reactions to replete NAD+ stores in the body [20]. An extracellular conversion of NMN to NR by CD73, a cell surface enzyme, represents another mechanism by which the intracellular NAD+ content is maintained [21].

Cellular levels of NAD+ are also regulated by the enzyme nicotinamide N-methyltransferase (NNMT). When NNMT methylates nicotinamide, creating methylnicotinamide (MNT), the amount of free nicotinamide is reduced and, therefore, not available for conversion into NAD+ through the NAD+ salvage pathway. NNMT and MNT have been associated with obesity and diabetes mellitus type two [22]. However, NNMT also has been shown to stabilize SIRT1, thereby exerting beneficial metabolic effects and protecting against oxidative stress-induced cellular injury [23,24]. Methylnicotinamide has also been shown to increase lifespan [25]. However, many NNMT inhibitors have been developed, and promising data establish a potential use for these in the treatment of pathologic states including, but not limited to, cancer, obesity, metabolic disorders, and alcohol-related fatty liver disease [22,26–29]. Taken together, the interaction between NNMT, MNT, and the pathways they help regulate plays a significant role in NAD+ homeostasis and, thus, the complex disease states that undoubtedly influence the aging process.

2.1. NAD+, Sirtuins and Longevity-Promoting Pathway

Disruption of proper NAD+ and loss of protective sirtuin activity have emerged as the prime targets for NAD+-based interventions [30]. Administration of NAD+ precursors, namely NR and NMN, has been shown to alleviate age-related NAD+ pathology, particularly in the context of age-related diseases [13,31,32]. Aging has been associated with a decreased NAD+/NADH ratio in human plasma through the deterioration of NAD+ stores, rather than an increase in NADH [12]. Replenishment of NAD+ rescues mitochondrial regulatory function from NAD+ induced pseudohypoxic mitochondrial stress during aging [14].

SIRT1, a member of a protein family that is involved in the cellular response to various stressors, has been shown to be implicated in longevity, but experiments and analysis have yielded mixed and context-dependent results. Nonetheless, high-level athletes exhibit higher telomere length and lowered insulin resistance, correlating with higher levels of SIRT1 expression [33]. The beneficial activity of SIRT1 may rely on the deacetylation and subsequent activation of the Forkhead transcription factors FoxO and PGC1α [34,35]. FoxOs is a transcription factor involved in stress resistance, cell cycle arrest, apoptosis, and tumor suppression. Activation of FoxOs has been linked with longevity in worms and flies [36,37]. Important processes, including growth, development, metabolism, reproduction, and longevity, are regulated through insulin/insulin-like growth factor signaling (IIS) working through the activity of FoxO. This pathway has been shown to extend neuronal activity and longevity under low IIS conditions [38,39]. PGC1α exerts its influence on mitochondrial biogenesis, where deficits are apparent in metabolic disease states. Overexpression of PGC1α has shown improved insulin sensitivity in muscle [40–42]. AMPK, involved in energy expenditure, also exhibits similarity to and bi-directional innervation from SIRT1

and further inhibits mTOR, an inhibitory process that has also been linked to longevity. It further activates SIRT1 by increasing available NAD+ stores [43]. Furthermore, nuclear factor κB (NF-κB) signaling, a process involved in innate immunity, can be inhibited by SIRT1 activity to reduce prolonged inflammatory signaling [44]. Depending on NAD+ stores in the body, SIRT1 activity arises as an interesting target in age-related pathway manipulation to promote longevity (Figure 1) [31,45–47]. The importance of maintaining an adequate NAD+ level for optimal SIRT1 activity during aging may be a key factor in regulating longevity.
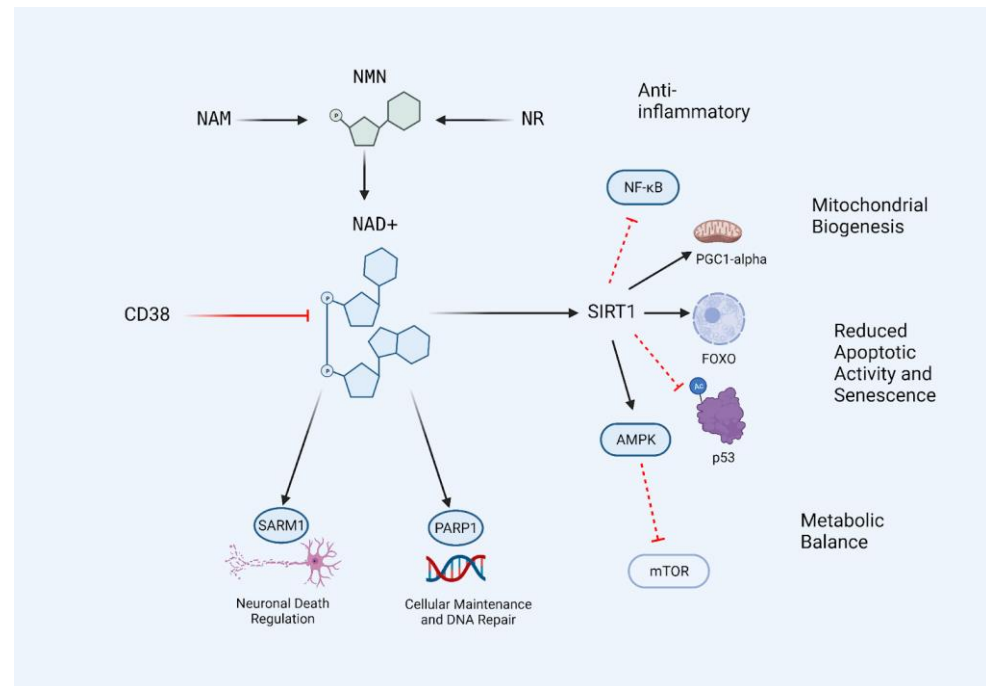

Figure 1. The CD38/NAD+/SIRT1 Axis. NAD+ levels in the body can be influenced by the supplementation of precursors nicotinamide (NAM), nicotinamide riboside (NR), and nicotinamide mononucleotide (NMN). NAD+ levels decrease with age and are further metabolized by the activation of SIRT1, PARP1, SARM1, and CD38. Restoring NAD+ levels allows for an increase in SIRT1 activity due to increased substrate availability, resulting in the inhibition of age-promoting pathways and activation of adaptive and protective transcription factors and processes. The central lineage may be described as the CD38/NAD+/SIRT1 axis, and targeting this access with nutraceutical interventions may prevent the age-related decline of NAD+ levels in the body. Black lines indicate conversion or activation. Red lines indicate inhibitors or destroyers of the indicated target.

2.2. NAD+ and Circadian Rhythm

NAD+ stores in the body are extremely crucial for the programming of the circadian metabolic clock. Older mice with less abundant NAD+ experienced prolonged repression of CLOCK/BMAL1 transcription compared to younger mice with more abundant NAD+, resulting in disrupted and dampened mitochondrial and transcriptional oscillation [48]. NAD+ supplementation and restoration in circadian mutant mice have also been shown to re-establish proper respiratory oscillations and circadian metabolic regulation, particularly through SIRT3 regulatory activity [49]. An abundant reserve of NAD+ and proper sirtuin activation are crucial for maintaining the integrity of the various endogenous clocks, and supplementation with NAD+ precursors may potentially alleviate any age-related perturbations in these circadian processes [50]. NAD+ deficiencies are observed in these numerous age-related diseases, and NAD+-based interventions are currently underway in an attempt to ameliorate this common conundrum these diseases share [51–54].

2.3. CD38 and the Decline of NAD+

Adequate intracellular levels of NAD+ are crucial for the proper function of several biological processes, including mitochondria metabolism [55]. In mitochondrial disease models, PARP inhibition or NR supplementation ameliorated deficits in metabolism and exercise capacity [56]. Strikingly, a cell surface glycoprotein, cluster of differentiation 38 (CD38), also acts as one of the main consumers of NAD+ stores in the body and is involved in immune activation and inflammatory signaling [57]. The NAD+ degrading enzyme CD38 has also been shown to regulate SIRT1 activity along with NAD+ availability [58]. CD38 expression increases during aging and CD38 knockout models exhibit heightened NAD+ levels and protection against obesity, metabolic disorders, and cancer progression [59], illuminating CD38′s role in increasing the risk of developing age-related metabolic diseases [60]. It has been suggested that CD38 deficiency may result in susceptibility to autoimmunity and decreased immune integrity, but further studies in humans are required to analyze the effects of insufficient CD38 [61]. Nevertheless, reducing chronic inflammation is an aim of healthy aging; inflammatory profiles of senescent cells are associated with heightened CD38 activation and resulting NAD+ degradation through compounding aging and inflammation processes [62]. Interestingly, this decline in NAD+ and increase in CD38 with age can potentially be attributed to another age-related phenomenon: the accumulation of senescent cells, cells that exhibit permanent growth arrest. Inflammation and aging, or "inflammaging," describes the build-up of inflammatory signaling compounds due to cellular senescence and the expression of CD38 as a senescent cellular inflammatory profile. Senescent cells, through their senescence-associated secretory profiles (SASP) of a milieu of IL-6, TNF-α, and CXCL 1 and 2, induce CD38 expression in M1 macrophages, causing age-related NAD+ decline in non-senescent phenotypic populations. These age-related senescent cells have also been shown to accumulate in hepatic and visceral white adipose tissue, and the resulting SASP induces pro-inflammatory M1 macrophage proliferation and enhanced CD38 expression [63–65]. The significant interplay between SASP, CD38, and NAD+ levels reveals an interventional target to further enhance available NAD+ in the body, particularly through strong inhibition of CD38 activity or expression well into the aging process to prevent any age-related biological deficits.

2.4. NMN as an NAD+ Boosting Therapeutic

When using an interventional method to target declining NAD+ levels, the bioavailability and proper uptake of the proposed supplement must be analyzed. NAD+ precursors, such as NR, NMN, and NAM, have been implicated in the adequate uptake and subsequent biosynthesis of NAD+ [66,67]. For instance, NAD3®, an NR-based dietary supplement, was shown in a clinical trial in older adults to significantly improve NAD+ levels as well as improve blood lipid profiles by decreasing LDL:HDL cholesterol ratios [68]. Another meta-analysis revealed that NAD+ boosting mechanisms are successful in decreasing total cholesterol, triglyceride, and the LDL:HDL ratio [69]. Moreover, NMN in particular serves as a molecule of interest in the context of NAD+ support, mainly due to the fact that only one conversion step is required to reach NAD+, compared to more required steps for other precursors. NMN supplementation has had numerous benefits, ranging from cardiovascular to neurodegenerative disease contexts, and proper uptake and resulting biosynthesis of NAD+ is observed in varying tissue types [70–72]. A successful increase in intracellular NAD+/NADH ratio was evident in a clinical trial in adults aged 40 to 65 years over 60 days at a dosage of 300 mg daily, providing key safety and efficacy data to support the use of NMN Uthever™ as a valid interventional approach to increase NAD+ in a safe manner [73]. NMN supplementation has also been shown to target one of the hallmarks of aging, telomere attrition. Short term supplementation of NMN in pre-aging mice and humans, ages 45 to 60 years, showed a significant increase in telomere length in peripheral blood mononuclear cells, as well as altered fecal microbiota benefiting immune and metabolic pathways [74], where specific administration in mice has been shown to increase the ratio of beneficial to harmful gut bacteria strains [75]. NMN supplementation has also

been shown to enhance the miRNA vascular expression profile in aged mice [76]. Respiratory parameters and aerobic capacity can also be enhanced through six weeks of NMN supplementation in adult runners, likely attributed to the enhanced oxygen utilization occurring in skeletal muscle as a result of increased NAD+ availability [77]. Furthermore, daily NMN supplementation at 250 mg was shown to be well tolerated, efficacious when increasing the NAD+/NADH ratio, and improved muscle function in aged but otherwise healthy men [78]. NMN supplementation has also been implicated in improving circadian rhythm patterns in older adults, where 250 mg daily for 12 weeks led to improvements in sleep quality, as measured through PSQI, fatigue, and physical performance [79].

Nevertheless, caution is warranted, as more recent data showed that in response to NR administration, increased rates of brain metastasis were observed after intracardial injection of triple-negative breast cancer (TNBC) cells in 9 out of 11 NR-treated mice compared to 3 out of 12 mice treated controls, proposing the increased risk of cancer metastasis in response to NR administration [80]. A contrasting study, however, showed no adverse effects on rats after administration of NMN at doses of 375, 750, and 150 mg/kg/d, with a maximum acute dose identified at 2666 mg/kg/d, enhancing the safety profile of NMN supplementation [81] To understand if there are differences in the safety of NAD+ precursor use, additional research is needed. To help to understand the translatability and mechanistic actions described in animal models, several clinical trials are ongoing (Table 1).

Table 1. Various clinical trials (completed and ongoing) detailing the use of NMN and other NMN derivates to improve health, metabolic markers, and disease parameters.

Clinical Trials	Compound of Interest
Nicotinamide Mononucleotide Increases Muscle Insulin Sensitivity in Prediabetic Women [82]	NMN
Effect of Oral Administration of Nicotinamide Mononucleotide on Clinical Parameters and Nicotinamide Metabolite Levels in Healthy Japanese Men [83]	NMN
Nicotinamide Mononucleotide Supplementation Enhances Aerobic Capacity in Amateur Runners: a randomized, double-blind study [77]	NMN
Effect of 12-Week Intake of Nicotinamide Mononucleotide on Sleep Quality, Fatigue, and Physical Performance in Older Japanese Adults: a randomized, double-blind placebo-controlled study [79]	NMN
Safety Evaluation of Beta-nicotinamide Mononucleotide Oral Administration in Healthy Adult Men and Women [84]	NMN
The Efficacy and Safety of Beta-nicotinamide Mononucleotide (NMN) Supplementation in Healthy Middle-aged Adults: a randomized, multicenter, double-blind, placebo-controlled, parallel-group dose-dependent clinical trial [85]	NMN
A Multicenter, Randomized, Double-Blind, Parallel Design, Placebo-Controlled Study to Evaluate the Efficacy and Safety of Uthever (NMN Supplement), an Orally Administered Supplementation, in Middle-Aged and Older Adults [73]	NMN
MIB-626, an Oral Formulation of a Microcrystalline Unique Polymorph of Beta-Nicotinamide Mononucleotide, Increases Circulating Nicotinamide Adenine Dinucleotide and its Metabolome in Middle-Aged and Older Adults [86]	MIB-626
Phase 2a MIB-626 vs. Placebo COVID-19 (NCT05038488)	MIB-626
Effect of Oral NAD+ Precursors Administration on Blood NAD+ Concentration in Healthy Adults (NICO) (NCT05517122)	NAM, NR, and NMN
Effect of NMN Supplementation on Organ System Biology (VAN) (NCT04571008)	NMN
Pharmacodynamics and Tolerance of Nicotinamide Mononucleotide (NMN, 400mg/Day) in Healthy Adults (NCT04862338)	NMN
Study to Evaluate the Effect of Nicotinamide Mononucleotide (NMN) As an Adjuvant to Standard of Care (SOC) On Fatigue Associated with Covid-19 Infection (NCT05175768)	NMN
Nicotinamide Mononucleotide in Hypertensive Patients (NCT04903210)	NMN
Safety and Pharmacokinetics of Nicotinamide Mononucleotide (NMN) in Healthy Adults (NCT04910061)	NMN
Effect of NMN (Nicotinamide Mononucleotide) on Polycystic Ovary Syndrome (NMN) (NCT05305677)	NMN
Effect of NMN (Nicotinamide Mononucleotide) on Diminished Ovarian Reserve (Including Premature Ovarian Insufficiency) (NCT05485610)	NMN
Effect of NMN on Muscle Recovery and Physical Capacity in Healthy Volunteers with Moderate Physical Activity (NCT04664361)	NMN

2.5. A Combination Approach for Restoring Youthful NAD+ Levels during Aging

As the interest in geroprotective trends grows, the momentum of clinical trials utilizing NAD+ boosting molecules is increasing. However, proper metrics evaluating the benefits and efficacy of intervention of NAD+ boosting supplements must be further standardized to offer significant and informative comparative effects of geroprotective interventions. NAD+ precursors have shown promising effects in human trials (Table 1), but with the involvement of CD38 and SIRT1 pathways, many potential interactions exist to further amplify the NAD+ boosting effects of orthomolecular interventions. SIRT1 activation, NAD+ synergy, CD38 inhibition, and methylation support are all important factors when considering maximizing the potential of NAD+ precursor administration. The synergistic effects and health benefits of the compounds involved in these three processes will be further discussed to create a foundation to pave the way for potentiating supplement formulations for maximal NAD+ boosting effects. One approach to maximize NAD+ boosting capacity would be to supplement safe levels of NMN with other geroprotectors and nutraceuticals that will further enhance endogenous NAD+ levels, ensuring the restoration of physiological levels of NAD+. Therefore, we summarized below different geroprotectors that enhance endogenous NAD+ levels but also have their own longevity benefits. For a selection of the following compounds, synergistic health effects of NMN and a geroprotector have been reported, suggesting the benefit of this combinatorial approach.

3. Potential Synergistic Nutraceutical Interventions

3.1. Stilbenes: Resveratrol and Pterostilbene

Some of the most extensively studied nutraceuticals are the non-flavonoid phenolic stilbene compounds resveratrol and pterostilbene [87] (Figure 2). These compounds are naturally occurring in grapes and berries and possess a plethora of anti-inflammatory and antioxidant characteristics, and even combat many age-related disorders such as diabetes and cancer [88]. Clinical analyses have identified these stilbenes as safe and bioavailable substances with doses of resveratrol up to 5 grams and pterostilbene to 250 mg [89,90].

However, resveratrol and pterostilbene have been shown to increase lifespan only in specific preclinical models, were context-dependent, showed mixed results, and are debated [91]. Pterostilbene has been shown to improve antioxidant function and increase longevity by activating longevity-associated genes, namely SIRT2, and has varying effects in male and female *Drosophila* models with the same longevity-inducing results but differing mechanisms [92]. Pterostilbene exhibits an 80% increased bioavailability, compared to that of resveratrol at 20%, and possesses many of the same beneficial biologically modulating effects of resveratrol. In the context of many disease states and aging, pterostilbene functions by upregulating endogenous antioxidant enzymes such as superoxide dismutase (SOD) and glutathione reductase (GR) [93]. The bioavailability of pterostilbene is an important factor in modulating SIRT1, as resveratrol has a much lower bioavailability and is metabolized quickly in comparison. Co-administration of resveratrol and pterostilbene may serve to maximize total bioavailability and maximize the benefits of each [94]. In response to particulate matter-induced oxidative damage, pterostilbene is able to mitigate the inflammatory response by decreasing mediators such as cyclooxygenase 2 (COX2), mitogen-activated protein kinase (MAPK), and the collagen-degrading matrix metalloproteinase (MMP)-9 expression, preventing the keratinocyte aging processes [95–98]. Resveratrol provides skin health benefits through its anti-angiogenic and wound-healing properties [99,100].

While resveratrol may also have numerous other targets, experimental models have shown a close association with the SIRT1 pathway when investigating resveratrol dosage and supplementation [101,102]. Resveratrol has exhibited many health benefits but yielded mixed results when used for the treatment of disease specifically, and the role of resveratrol in the direct activation of SIRT1 has been heavily questioned. Resveratrol seems to be at least indirectly involved in the activation of SIRT1 to induce the beneficial effects of caloric restriction, which has been shown numerous times to induce lifespan extension [103]. The activation of SIRT1 along with AMPK and PGC1α in response to resveratrol is implicated,

but despite this lack of clarity in interventional disease trials, resveratrol might still be considered an age-related metabolic modulator [104].
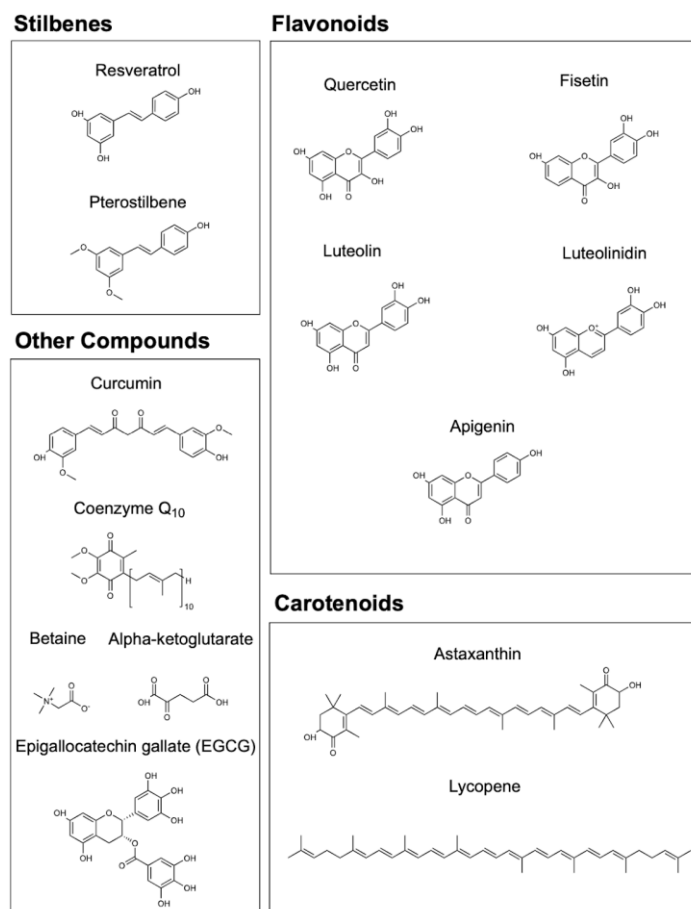

Figure 2. Chemical structures of the aforementioned nutraceutical compounds are organized into their respective categories.

Resveratrol may also enhance neuroprotection by reducing neuroinflammation, thereby improving cognition and neuro-metabolic protection and decreasing the risk of age-related neurodegeneration. Resveratrol also possesses the ability to improve oxidative stress in the vasculature by reducing superoxide generation and increasing the antioxidant response of endothelial cells, thereby dissipating the age-related risk of cardiovascular disease, also initiated through the SIRT1-involved pathway. Anti-metastatic and anti-cancer properties are notoriously associated with resveratrol, as it helps suppress tumor proliferation and uncontrolled cellular division [105]. Interestingly, resveratrol has the ability to mimic caloric restriction (CR) in a similar fashion to that associated with the SIRT1 pathway to extend longevity. This occurs through several mechanisms. First, increasing the activity of telomerase and FoxO3a, a transcription factor associated with increased longevity. It also occurs through active regulation of Sirt1 (AROS). Finally, increasing Hu antigen R (HuR) and decreasing p53 can prevent cellular senescence but inhibit tumor suppression [106–108]. It has also demonstrated the ability to improve biological parameters of kidney aging [109] and improves arterial aging by reducing media thickness, inflammation, fibrosis, and oxidative stress through increasing SIRT1, AMPK, and PGC1α, SOD1, and SOD2 expression, as well as decreasing NADH oxidase 2 and 4 [110,111].

At low concentrations, resveratrol can inhibit cellular senescence; however, at higher concentrations, it exhibits complete senolytic activity [112]. Resveratrol can even exert an influence on reproductive aging, as seen in the postovulatory oocytes of mice. Rates of fertilization and blastocyst size increased in response to resveratrol administration, as well

as upregulation of FoxO3a [113,114]. The cognitive effects of stilbene administration have gained more of a solid foundation in recent years, supporting its benefits in supplementation [115,116]. Cognitive impairment due to aging was attenuated by improved cognitive performance in older mice and decreasing levels of inflammatory mediators such as IL-1β and TNF-α in response to resveratrol [117], and aged mice that received resveratrol showed improvements in cerebrovascularity [118]. In response to a high-fat diet in rats, resveratrol or alpha-ketoglutarate (aKG) improved brain and metabolic aging profiles, notably through inflammatory response genes [119]. Witte et al., 2014, analyzed resveratrol administration at 200 mg daily in older adults to affect memory performance, hippocampal connectivity, and glucose metabolism. The results were that memory retention increased in the interventional group, along with increased functional connectivity in the frontal, parietal, and occipital regions of the hippocampus. Hemoglobin A1c (HB1Ac) and body fat decreased, implying lowered blood sugar, and the satiety hormone, leptin, increased. Both changes correlated with increased memory retention and increased functional connectivity. These changes were consistent with the benefits reported in previous studies, further demonstrating enhanced cognitive function and improved metabolic profiles through stilbene supplementation [120,121]. In the context of neurodegeneration and Alzheimer's disease patients with mild impairment, resveratrol treatment resulted in a decrease in neuroinflammatory profile and MMP9 expression, but further studies are still required to validate the mechanism and efficacy in neurodegenerative disorders [122].

In the treatment of diabetes, resveratrol has been shown to improve oxidative and inflammatory microRNA profiles and reduce insulin resistance in diabetic humans [123]. Interestingly, resveratrol administration has been quite beneficial in ameliorating post-menopausal symptoms in older women. The risk of osteoporosis and improvements in non-osteoporotic post-menopause-related bone density were reported in response to 75 mg of daily resveratrol administration [124]. A 24-month crossover study in 125 postmenopausal women showed marked improvements in cognition, cerebrovascular function, and insulin sensitivity [125], and sustained cognitive, cardiovascular, and cerebrovascular benefits were reported over a 12-month study period in response to the same daily dosage, 75 mg, of resveratrol [126]. These results strongly implicate resveratrol ameliorating age-related dysfunction in post-menopausal women as a preventative measure.

Resveratrol combined with the senolytic flavonoid quercetin has also been shown to improve aging panels of the kidney in high glucose conditions by reducing the glycation precursor, methylglyoxal, and increasing senescence marker protein 30 (SMP30) expression [127]. Rat heart tissue has also shown improved inflammatory, apoptotic, and oxidative panels in response to resveratrol administration, particularly through the reduction of the inflammatory markers TNF-α and IL-gamma [128]. In particular, Ghanim et al. demonstrated that in response to a high-fat and high-carbohydrate meal, 100 mg of resveratrol and 75 mg of grape skin polyphenols were able to upregulate the binding activity of oxidative stress-response transcription factor nuclear factor erythroid 2-related factor (Nrf2) to detoxification target genes NAD(P)H Quinone Dehydrogenase 1 (NQO-1) and glutathione-S-transferase pi 1 (GST-pi1), as well as ameliorate the effects of inflammatory mediators such as the cluster of differentiation 14 (CD14), IL-1β, and Toll-like receptor 4 (TLR4), potentiating their anti-diabetic, anti-inflammatory, and anti-atherosclerosis effects [102,129]. Induced cartilage stiffening in response to ribose administration can also further be ameliorated by resveratrol as a result of decreased glycation and advanced glycation end products (AGE) because of the competitive collagen II binding activity of resveratrol and curcumin [130]. Resveratrol can further improve insulin sensitivity and skeletal aging through the SIRT1 pathway by reducing FoxO1-induced transactivation of pyruvate dehydrogenase kinase 4 (PDK4), increasing muscular metabolism integrity [131,132].

Furthermore, resveratrol and pterostilbene's effects might be heightened when used in combination with NMN supplementation to produce optimal results as a cleverly tailored nutraceutical orthomolecular approach to delaying or even reversing the signs of aging [133]. Age- and risk-related biological markers for certain diseases can also be

monitored in response to resveratrol and pterostilbene administration [134,135]. Coadministration of NMN and resveratrol resulted in increased NAD+ levels in the heart and skeletal muscle compared to NMN alone. NMN and ginsenoside coadministration resulted in higher NAD+ levels compared to NMN alone in the brain, heart, kidney, and lung tissues, further supporting the dual supplementation approach to maximize NAD+ levels in the body [136]. Resveratrol also retains the ability to activate NMNAT1, the NAD+ synthetic enzyme, to increase NAD+ levels by up to five times, providing a large substrate store for SIRT1 activation to promote longevity-enhancing pathways [137]. NR and pterostilbene co-administration additionally exhibit a two-dose, dose-dependent increase in NAD+ levels in acute kidney injury patients [138], and repeated doses of NR and pterostilbene also exhibit an increase in NAD+ levels of 40% and 90% (double dose) in humans with a consistent increase over the 8-week trial duration [139]. Resveratrol and quercetin are also able to promote cell protection and NAD+ levels along with NAD+ precursors by potentiating the cell rescue effects of said precursors in response to DNA damage [140]. Thus, by supporting the transient supplementation of NMN, stilbene combinations may maximize the potential for this longevity-based orthomolecular approach.

3.2. CoQ10

Coenzyme Q10 (CoQ10; Figure 2), or ubiquinol (the oxidized form of ubiquinone), participates in the electron transport chain in the mitochondria, and low levels of CoQ10 are associated with diseases such as neurodegenerative disorders, diabetes, cancer, fibrosis, and cardiovascular diseases [141]. Supplementation with CoQ10 in the context of disease states aims to re-establish antioxidant activity to ameliorate homeostatic disruptions [142]. CoQ10 has protective cardiovascular qualities and can result in improvements in hyperglycemia, hypertension, oxidative stress, and risk of cardiac events [143]. Endogenous biosynthesis of CoQ10 sharply decreases with aging, and enhanced longevity can be seen with higher mitochondrial levels of CoQ10. Skeletal muscle integrity in elderly individuals is associated with higher plasma CoQ10 content, and inflammatory factors such as TNF-α, IL-6, and C-reactive protein (CRP) levels can be reduced as a result of supplementation [144]. CoQ10 is also required for lipid integrity and protection of LDL oxidation against atherosclerosis [145]. Replenishing the age-related decline in levels of CoQ10 is imperative to reduce the risk of age-related diseases and lower the burden of age-inducing oxidative stress through nutraceutical supplementation [146]. CoQ10 supplementation, along with dietary interventions in elderly men and women, resulted in improved metabolic profiles, reducing metabolic and cardiovascular risk [147].

CoQ10 and NAD+ supplementation also portray a degree of synergy when analyzing maximum heart rate after exercise in chronic fatigue syndrome (CFS). Maximum heart rate was decreased, and reports of fatigue improved [148]. CFS shares some characteristics with aging and age-deregulated pathways, such as inflammation and oxidative stress, and CoQ10 and NADH supplementation were further shown to improve fatigue and levels of NAD+/NADH, CoQ10, ATP, citrate synthase, and lipoperoxides [149].

The antioxidant, anti-inflammatory, and age-related effects of CoQ10 make it a highly appealing supplement to include in the comprehensive orthomolecular approach to battling the biological process of aging, particularly in the context of supporting NAD+ levels. Further studies are required to show, for instance, the beneficial synergistic effects of NAD+ precursor supplementations in combination with CoQ10.

3.3. Betaine

Trimethylglycine, known as betaine, was originally derived from the beetroot plant (Figure 2). It has osmoprotectant and anti-inflammatory properties. Betaine, along with methionine and choline, is one of the primary methyl group donors involved in DNA methylation, and DNA methylation rates have been directly correlated with available access to methyl donors [150]. Betaine also suppresses numerous inflammatory expression profiles, such as TNF-α, COX2, and NF-kB activity [151]. The effects of betaine are implicated in

postponing aging-related pathologies by promoting optimal lipid and glucose metabolism, inhibiting inflammatory transcription processes, and reducing cellular ER stress [152].

Interestingly, betaine and choline levels are influenced by the degradation of the NAD+ precursors, namely NAM, which consumes more betaine than choline. This potentially compromises the availability of a healthy pool of methyl donors [153]. The high burden of NAM degradation on betaine levels and epigenetic changes warrants adequate supplementation of methyl donors during concomitant NAD+ precursor administration, particularly NAM [154], but the effects of NMN or NAD+ conversion on methylation levels have not been explored. Concurrent supplementation of NMN, NAD+, or other precursors alongside betaine may prevent the proposed decline in betaine levels, maintaining proper methylation health and function.

3.4. Flavonoids: Quercetin, Fisetin, Luteolin/Luteolinidin, and Apigenin

Numerous flavonoids have also been shown to have a host of health benefits along with strong senolytic activity, such as fisetin, quercetin, luteolin/luteolinidin, and apigenin.

Fisetin and quercetin are potent anti-cancer agents and have been shown to have anti-tumor activity through calcium-induced tumor apoptosis along with improving cancer-related inflammatory profiles [155].

Fisetin has been shown to be a potent senolytic in older and progeroid models of mice, as well as in murine and human adipose tissue (Figure 2). Administration of fisetin improved lifespan and tissue homeostasis in mice [156]. Senolytic effect is a crucial characteristic of an interventional dietary supplement, and fisetin has been shown to perform as one of the strongest among numerous other flavonoids and compounds. The safety and efficacy of fisetin are currently being investigated in reducing inflammation and improving walking speed in frail elderly individuals in Phase 2 randomized clinical trials (NCT03675724, NCT03430037). Interestingly, fisetin also portrays a degree of interrelatedness to NAD+/NADH age-related pathology through its activation of SIRT1 by inducing mitochondrial fragmentation in a similar fashion to treatment with NAM and an increased NAD+/NADH ratio [157]. These findings point to the possibility that fisetin may have geroprotective effects in the context of the NAD+/SIRT1/CD38 pathway, but further experiments must be conducted to see measurable and explicit effects on the biochemical longevity axis.

Quercetin, another flavonoid with striking structural similarity to fisetin (Figure 2), is also considered a senolytic with many benefits in the realm of cardiovascular disease, neurodegeneration, inflammation, oxidative stress, cancer, and diabetes. Quercetin has been shown to be a geroprotective agent for in vitro models of premature aging [158,159]. Quercetin also exhibits attractive properties as a supplemental ingredient in the NAD+ boosting approach. Along with the previously mentioned apigenin, quercetin has been shown in the same experiment to be a potent CD38 inhibitor to maintain the integrity of NAD+ stores and act as a protective agent against metabolic disorders [160]. Quercetin can potently initiate SIRT1's intermediary action to result in anti-inflammatory and antioxidant effects in diet-induced atherosclerosis rat models [161]. NAD+/NADH balance is also further affected by quercetin's ability to oxidize NADH and decrease the NADH/NAD+ ratio, where larger concentrations of the flavonoid stimulated the Krebs cycle, likely due to an increase in available NAD+ [162]. Nevertheless, quercetin earns a role in the modulation of the NAD+/SIRT1/CD38 axis due to its effects on altering the NAD+/NADH ratio, SIRT1 activation, and CD38 inhibition.

Luteolin, another flavonoid with similar health benefits as other geroprotectors, also has strong anti-inflammatory capabilities in relation to skin aging, skin disease, skin cancer, and cognition [163,164] (Figure 2). Luteolin is implicated in CD38 mechanisms along with apigenin and quercetin (and other compounds, such as luteolinidin and kuromanin) and has been shown to be a potent inhibitor of CD38, ultimately leading to an increase in available NAD+ [165]. Luteolinidin also exhibits myocardial protective attributes by stabilizing NADPH and mitigating the ischemic injury-induced upregulation of CD38 [166].

Flavonoids may have further longevity-promoting effects through the clearance of cellular senescence during concurrent administration of NAD+ supporting compounds, and further compounds may be tested on their activity on NAD+/NADH ratios and sirtuin activation [167].

The flavonoid apigenin is derived from plants, such as parsley and chamomile, and possesses highly anti-inflammatory, antioxidant, and anti-carcinogenic properties (Figure 2). It can reduce COX2, IL6, and TNF-α in response to bacterial toxins and lipopolysaccharide (LPS) [168]. Upregulation of antioxidant enzymes such as SOD, glutathione peroxidase (GPX), and glutathione reductase (GR) is also evident in response to lower dose apigenin administration [169]. Apigenin achieves its anti-carcinogenic activity by downregulating key factors in cancer pathways and sensitizing tumor cells to chemotherapeutic interventions [170]. In the context of esophageal cancer, IL-6 transcription is inhibited in response to apigenin as an intervention [171], and apigenin also exhibits anti-leukemic activity in vivo and in human cells by inducing apoptosis [172]. Apigenin can further attenuate metabolic complications leading to the risk and characteristics of metabolic disorders, such as metabolic syndrome, atherosclerosis, and diabetes [173]. Apigenin reduces the expression of COX2 and can also produce anti-inflammatory effects in the context of cardio-vascular disease, neurodegeneration, diabetes, and cancer [174]. Apigenin supplementation has anti-obesity effects, with marked reductions in visceral adipose tissue in high-fat-fed mice, and can preserve skeletal muscle in obese mouse models of obesity-induced skeletal muscle atrophy [175,176]. Apigenin also possesses the ability to ameliorate deleterious age-related changes in vascular endothelial function and structure due to oxidative stress [177]. The anti-inflammatory and anti-cancer properties of apigenin make it appealing to utilize in the context of healthy aging and geroprotection [178].

The most notable property of apigenin in the context of NAD+ supplementation is the involvement with the SIRT1 NAD+ and CD38 axis. Apigenin administration increases the activation ratio of SIRT1, NAD+, NAD+/NADH and strongly inhibits CD38, thereby enhancing endogenous NAD+ levels. The resulting increase in SIRT1 reduced cellular senescence as a result of oxidative stress [179]. Apigenin is a potent inhibitor of CD38, and knockout models have shown resistance to metabolic disorders and exhibit increased NAD+ levels. Inhibition of CD38 in obesity models led to an enhanced metabolic profile, increased NAD+ levels, and decreased protein acetylation, possibly due to enhanced SIRT1 activity due to higher NAD+ availability [160]. CD38 inhibition through apigenin administration additionally restores mitochondrial function in response to oxidative stress and increases the NAD+/NADH ratio and SIRT3 activation [180]. Apigenin serves as a crucial component in the comprehensive approach to restoring age-related depletion of NAD+ levels through its strong inhibition of CD38, the degrading enzyme of NAD+. Through its CD38 inhibitory activity, apigenin becomes a valuable interventional agent when maximizing the effects of increasing NAD+ availability in the body through NMN supplementation.

3.5. Carotenoids: Astaxanthin and Lycopene

Astaxanthin is a potent antioxidant carotenoid compound that offers an array of health benefits by mitigating reactive oxygen species (ROS) and supporting mitochondrial integrity [181] (Figure 2). Unsurprisingly, astaxanthin is highly efficacious in utilizing SIRT1 to achieve longevity-promoting effects in many experimental models. Astaxanthin was shown in vivo to soothe oxidative stress in response to brain injury by upregulating the expression of Nrf2 and SIRT1 while concurrently reducing the expression of pro-apoptotic factors, potentially reducing the risk of neuronal death events post-injury [182]. The effects of a high-fat-induced diet on cardiac and fibrotic damage can also be ameliorated by astaxanthin through the upregulation of SIRT1, inhibition of inflammatory cellular mobility, and reduced incidence of collagen deposition leading to fibrosis post-injury [183,184]. Astaxanthin can additionally protect renal tissue post-injury through its upregulation of SIRT1 [185]. The most supportive finding of astaxanthin's ability to boost NAD+ availability was found by Zhang et al., where NMN, astaxanthin, and blood orange extract were utilized

to successfully increase NAD+ in aging zebrafish, exhibiting superiority in raising NAD+ levels compared to supplementation of just NR, NR and astaxanthin, and even NR and pterostilbene [186]. This study can serve as a basis for future studies of supplements to hone in on effective and precise dosages and synergistic combinations to maximize the effect of NAD+ boosting approaches in humans.

Lycopene, another red-pigmented carotenoid, also possesses strong antioxidant and anti-inflammatory characteristics (Figure 2). Oxidative stress and the intrinsic aging process can lead to plasma lycopene depletion and impaired absorption, and supplementation has been shown to improve physical performance, osteoporosis, and skin aging [187]. Interestingly, lycopene activates SIRT1 to impart muscle angiogenesis and reverse insulin resistance in models of age-related vascular decline [188]. In vivo and in vitro models of D-galactose-induced aging were ameliorated in a superiority trial with astounding results in response to NMN and lycopene combination therapy, with increases in SOD; GSH-px, GPX, CAT, T-AOC, and evidence of senolytic abilities. Nrf2 was upregulated, and in vivo models showed enhanced cognition compared to NMN alone [189].

3.6. Curcumin

Curcumin has earned a role as a potent senolytic, along with the previously mentioned flavonoids (Figure 2). It has been implicated in improving senescence in age-related pathologies and has even exhibited modulatory effects on longevity-related pathways, such as the mTOR and FoxO [190]. In the context of neurodegeneration, curcumin has been shown to upregulate SIRT1 and, in the cardiovascular context, AMPK [191]. Anti-cancer, migration, and anti-angiogenesis activity can be observed in response to curcumin administration in head and neck squamous cell carcinoma experimental cell models [192]. Six-week curcumin supplementation has been shown to improve antioxidant and aerobic capacity in human runners, along with an increase in SIRT3 [193]. The connection between curcumin and sirtuins is evident, but the utilization of curcumin along with NAD+ boosting molecules must be first demonstrated in future superiority trials involving combination therapies.

3.7. Alpha-Ketoglutarate

Alpha-ketoglutarate (aKG; Figure 2) is a crucial metabolic intermediate that is involved in the Krebs cycle and is implicated in the process of aging [194]. Along with other geroprotectors that inhibit the TOR pathway, AKG is able to additionally inhibit ATP synthase to extend the lifespan in *C. elegans* in a fashion similar to caloric restriction [195]. Offering metabolic and antioxidant benefits [196], aKG produces lifespan-extending benefits which are additionally apparent in mice, and recent pilot clinical trials in humans have shown that Rejuvant, a novel formulation of aKG, was successful in reducing the biological age of study participants [197,198]. However, data involving aKG and NAD+ interplay are lacking and must be explored further.

3.8. Epigallocatechin Gallate

The polyphenol epigallocatechin gallate (EGCG; Figure 2), found in green tea, has been touted to retain neuroprotective, antioxidant, and anti-inflammatory effects and is currently being analyzed for its role in ameliorating numerous different diseases [199]. It has been shown to increase rat lifespan in response to oxidative stress [200]. However, the role of EGCG in SIRT1 modulation is unclear, as certain studies see upregulation of SIRT1 in response to administration [201], but others observe downregulation, particularly in cancerous cells [202,203]. EGCG may behave differently on SIRT1 depending on the context and which longevity factors must be upregulated to mitigate oxidative stress, and the effects of EGCG on NAD+/NADH ratio must be further observed.

4. Conclusive Remarks

Having established the benefits of numerous ingredients involved in longevity-related pathways, a clearer picture emerges of the entangled molecular web of the NAD+/SIRT1/CD38

axis in the context of healthy aging supplementation (Figure 3). Many ingredients on their own show numerous benefits in the realm of many age-related disorders, such as neurodegeneration, cardiovascular and metabolic disease, diabetes, cancer, frailty, sarcopenia, hearing loss, osteoarthritis, osteoporosis, and ocular disorders [204]. CD38 inhibitors may become a focus for developing or screening new compounds, which may imply longevity induction in vivo and, ultimately, in humans. CD38-inhibitor 78c is one compound that has yet to enter human interventional trials but has shown marked improvements in longevity, physical performance, and metabolic profile in mice [205].
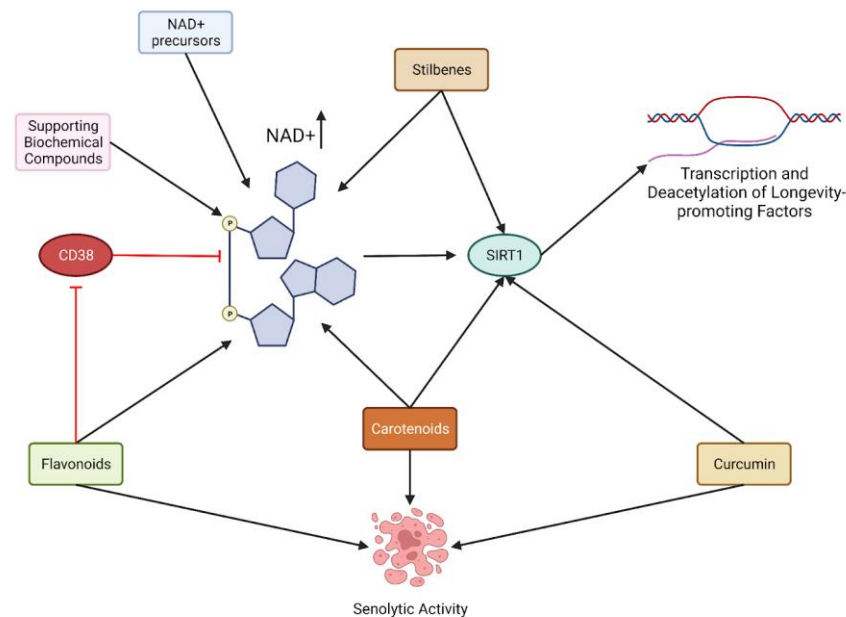
Figure 3. Hypothesized model of supplementing NAD+ precursors with other NAD+ enhancing geroprotectors. In addition to NAD+ precursors to raise NAD+ levels and enhance SIRT1 activity, stilbenes are able to support NAD+ levels and further activate SIRT1. Many flavonoids retain senolytic and CD38-inhibitory activity and can further innervate NAD+ stores. Curcumin and carotenoids retain similar properties in addition to SIRT1 activation. This interacting web of support may result in higher NAD+ stores than precursor-alone administration, producing longevity-promoting transcriptional benefits.

A beneficial direction may be to further look into other senolytics that ameliorate numerous hallmarks of aging. Further clarification of SIRT1 activity in relation to downstream effects implicated in longevity extensions shown in experimental models is required, particularly in humans, to determine the maximal effect of innervation through nutraceutical interventions. Testing superiority trials of NMN or NR in combination with other ingredients compared to NMN or NR alone, geroprotectors, senolytics alone, or other NAD+ precursor molecules will prove vital in maximizing the effects of each ingredient along with maximal NAD+ boosting potential. The potential of generated knowledge from further analyzing these relationships within the context of human aging may also offer insight into other pathways that may be implicated in longevity but may yield mixed results in the context of the fine balance for tumor suppression, senescence, and senolytic activity. Subsequently, avenues to target the pathology of aging and age-related diseases to increase healthspan may arise upon further exploration of other geroprotectors, particularly within clinical trials.

Due to the lack of superiority trials analyzing combination therapies and potential molecular synergies, a call for further RCTs is highly warranted in the field of healthy aging orthomolecular medicine and geroprotective nutrition. By screening for compounds that exhibit an effect on aging-related molecular pathways, innovative formulations of geroprotective ingredients can be achieved. Perhaps the maximum benefit can be derived

not simply from one compound but from an ingredient profile that incorporates multiple compounds that work together to achieve a synergistic effect. If a formulation can function to boost NAD+ levels and SIRT1 activity and/or reduce CD38 activity, this will optimize lifespan and healthspan via extension supplements. Further, the more pathways that are affected, the more broadly these supplements can be applied to have a positive impact on the health and aging of multiple bodily systems. This will expand the application of these supplements, thereby broadening the populations that they may benefit.

References

1. Niccoli, T.; Partridge, L. Ageing as a risk factor for disease. *Curr. Biol.* **2012**, *22*, R741–R752. [CrossRef] [PubMed]
2. Jose, V.; Consuelo, B.; Miquel, J. Theories of ageing. *IUBMB Life* **2007**, *59*, 249–254. [CrossRef]
3. Jin, K. Modern Biological Theories of Aging. *Aging Dis.* **2010**, *1*, 72–74.
4. Partridge, L. Intervening in ageing to prevent the diseases of ageing. *Trends Endocrinol. Metab.* **2014**, *25*, 555–557. [CrossRef] [PubMed]
5. Janson, M. Orthomolecular medicine: The therapeutic use of dietary supplements for anti-aging. *Clin. Interv. Aging* **2006**, *1*, 261–265. [CrossRef]
6. Borumand, M.; Sibilla, S. Daily consumption of the collagen supplement Pure Gold Collagen® reduces visible signs of aging. *Clin. Interv. Aging* **2014**, *9*, 1747–1758. [CrossRef]
7. Skovgaard, G.R.L.; Jensen, A.S.; Sigler, M.L. Effect of a novel dietary supplement on skin aging in post-menopausal women. *Eur. J. Clin. Nutr.* **2006**, *60*, 1201–1206. [CrossRef]
8. Stephens, T.J.; Sigler, M.L.; Hino, P.D.; Moigne, A.L.; Dispensa, L. A Randomized, Double-blind, Placebo-controlled Clinical Trial Evaluating an Oral Anti-aging Skin Care Supplement for Treating Photodamaged Skin. *J. Clin. Aesthet. Dermatol.* **2016**, *9*, 25–32.
9. Poddar, J.; Pradhan, M.; Ganguly, G.; Chakrabarti, S. Biochemical deficits and cognitive decline in brain aging: Intervention by dietary supplements. *J. Chem. Neuroanat.* **2018**, *95*, 70–80. [CrossRef]
10. Lemon, J.A.; Boreham, D.R.; Rollo, C.D. A Complex Dietary Supplement Extends Longevity of Mice. *J. Gerontol. Ser. A* **2005**, *60*, 275–279. [CrossRef]
11. Xie, X.; Gao, Y.; Zeng, M.; Wang, Y.; Wei, T.-F.; Lu, Y.-B.; Zhang, W.-P. Nicotinamide ribose ameliorates cognitive impairment of aged and Alzheimer's disease model mice. *Metab. Brain Dis.* **2019**, *34*, 353–366. [CrossRef]
12. Clement, J.; Wong, M.; Poljak, A.; Sachdev, P.; Braidy, N. The Plasma NAD+ Metabolome Is Dysregulated in "Normal" Aging. *Rejuvenation Res.* **2019**, *22*, 121–130. [CrossRef]
13. Fang, E.F.; Lautrup, S.; Hou, Y.; Demarest, T.G.; Croteau, D.L.; Mattson, M.P.; Bohr, V.A. NAD+ in Aging: Molecular Mechanisms and Translational Implications. *Trends Mol. Med.* **2017**, *23*, 899–916. [CrossRef]
14. Gomes, A.P.; Price, N.L.; Ling, A.J.Y.; Moslehi, J.J.; Montgomery, M.K.; Rajman, L.; White, J.P.; Teodoro, J.S.; Wrann, C.D.; Hubbard, B.P.; et al. Declining NAD+ induces a pseudohypoxic state disrupting nuclear-mitochondrial communication during aging. *Cell* **2013**, *155*, 1624–1638. [CrossRef]
15. Radenkovic, D.; Reason; Verdin, E. Clinical Evidence for Targeting NAD Therapeutically. *Pharmaceuticals* **2020**, *13*, 247. [CrossRef]

16. Liu, J.-K. Antiaging agents: Safe interventions to slow aging and healthy life span extension. *Nat. Prod. Bioprospect.* **2022**, *12*, 18. [CrossRef]

17. Schultz, M.B.; Sinclair, D.A. Why NAD+ Declines during Aging: It's Destroyed. *Cell Metab.* **2016**, *23*, 965–966. [CrossRef]

18. Ray Chaudhuri, A.; Nussenzweig, A. The multifaceted roles of PARP1 in DNA repair and chromatin remodelling. *Nat. Rev. Mol. Cell Biol.* **2017**, *18*, 610–621. [CrossRef]

19. Gerdts, J.; Brace, E.J.; Sasaki, Y.; DiAntonio, A.; Milbrandt, J. SARM1 activation triggers axon degeneration locally via NAD+ destruction. *Science* **2015**, *348*, 453. [CrossRef]

20. Katsyuba, E.; Romani, M.; Hofer, D.; Auwerx, J. NAD$^+$ homeostasis in health and disease. *Nat. Metab.* **2020**, *2*, 9–31. [CrossRef]

21. Mateuszuk, Ł.; Campagna, R.; Kutryb-Zając, B.; Kuś, K.; Słominska, E.M.; Smolenski, R.T.; Chlopicki, S. Reversal of endothelial dysfunction by nicotinamide mononucleotide via extracellular conversion to nicotinamide riboside. *Biochem. Pharmacol.* **2020**, *178*, 114019. [CrossRef] [PubMed]

22. Kannt, A.; Rajagopal, S.; Kadnur, S.V.; Suresh, J.; Bhamidipati, R.K.; Swaminathan, S.; Hallur, M.S.; Kristam, R.; Elvert, R.; Czech, J.; et al. A small molecule inhibitor of Nicotinamide N-methyltransferase for the treatment of metabolic disorders. *Sci. Rep.* **2018**, *8*, 3660. [CrossRef] [PubMed]

23. Campagna, R.; Mateuszuk, Ł.; Wojnar-Lason, K.; Kaczara, P.; Tworzydło, A.; Kij, A.; Bujok, R.; Mlynarski, J.; Wang, Y.; Sartini, D.; et al. Nicotinamide N-methyltransferase in endothelium protects against oxidant stress-induced endothelial injury. *Biochim. Biophys. Acta Mol. Cell Res.* **2021**, *1868*, 119082. [CrossRef] [PubMed]

24. Hong, S.; Moreno-Navarrete, J.M.; Wei, X.; Kikukawa, Y.; Tzameli, I.; Prasad, D.; Lee, Y.; Asara, J.M.; Fernandez-Real, J.M.; Maratos-Flier, E.; et al. Nicotinamide N-methyltransferase regulates hepatic nutrient metabolism through Sirt1 protein stabilization. *Nat. Med.* **2015**, *21*, 887–894. [CrossRef] [PubMed]

25. Schmeisser, K.; Mansfeld, J.; Kuhlow, D.; Weimer, S.; Priebe, S.; Heiland, I.; Birringer, M.; Groth, M.; Segref, A.; Kanfi, Y.; et al. Role of sirtuins in lifespan regulation is linked to methylation of nicotinamide. *Nat. Chem. Biol.* **2013**, *9*, 693–700. [CrossRef]

26. Van Haren, M.J.; Zhang, Y.; Thijssen, V.; Buijs, N.; Gao, Y.; Mateuszuk, L.; Fedak, F.A.; Kij, A.; Campagna, R.; Sartini, D.; et al. Macrocyclic peptides as allosteric inhibitors of nicotinamide N-methyltransferase (NNMT). *RSC Chem. Biol.* **2021**, *2*, 1546–1555. [CrossRef]

27. Van Haren, M.J.; Gao, Y.; Buijs, N.; Campagna, R.; Sartini, D.; Emanuelli, M.; Mateuszuk, L.; Kij, A.; Chlopicki, S.; Escudé Martinez de Castilla, P.; et al. Esterase-Sensitive Prodrugs of a Potent Bisubstrate Inhibitor of Nicotinamide N-Methyltransferase (NNMT) Display Cellular Activity. *Biomolecules* **2021**, *11*, 1357. [CrossRef]

28. Neelakantan, H.; Vance, V.; Wetzel, M.D.; Wang, H.-Y.L.; McHardy, S.F.; Finnerty, C.C.; Hommel, J.D.; Watowich, S.J. Selective and membrane-permeable small molecule inhibitors of nicotinamide N-methyltransferase reverse high fat diet-induced obesity in mice. *Biochem. Pharmacol.* **2018**, *147*, 141–152. [CrossRef]

29. Gao, Y.; van Haren, M.J.; Buijs, N.; Innocenti, P.; Zhang, Y.; Sartini, D.; Campagna, R.; Emanuelli, M.; Parsons, R.B.; Jespers, W.; et al. Potent Inhibition of Nicotinamide N-Methyltransferase by Alkene-Linked Bisubstrate Mimics Bearing Electron Deficient Aromatics. *J. Med. Chem.* **2021**, *64*, 12938–12963. [CrossRef]

30. Mendelsohn, A.R.; Larrick, J.W. The NAD$^+$/PARP1/SIRT1 Axis in Aging. *Rejuvenation Res.* **2017**, *20*, 244–247. [CrossRef]

31. Yaku, K.; Okabe, K.; Nakagawa, T. NAD metabolism: Implications in aging and longevity. *Ageing Res. Rev.* **2018**, *47*, 1–17. [CrossRef]

32. Chini, C.C.S.; Tarragó, M.G.; Chini, E.N. NAD and the aging process: Role in life, death and everything in between. *Mol. Cell. Endocrinol.* **2017**, *455*, 62. [CrossRef]

33. Aguiar, S.S.; Rosa, T.S.; Neves, R.V.P.; Leite, P.L.A.; Maciel, L.A.; Gutierrez, S.D.; Rosa, E.C.; Andrade, R.V.; Degens, H.; Korhonen, M.T.; et al. Telomere Length, SIRT1, and Insulin in Male Master Athletes: The Path to Healthy Longevity? *Int. J. Sport. Med.* **2022**, *43*, 29–33. [CrossRef]

34. Tang, B.L. Sirt1 and the Mitochondria. *Mol. Cells* **2016**, *39*, 87–95. [CrossRef]

35. Brunet, A.; Sweeney, L.B.; Sturgill, J.F.; Chua, K.F.; Greer, P.L.; Lin, Y.; Tran, H.; Ross, S.E.; Mostoslavsky, R.; Cohen, H.Y.; et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. *Science* **2004**, *303*, 2011–2015. [CrossRef]

36. Greer, E.L.; Brunet, A. FOXO transcription factors at the interface between longevity and tumor suppression. *Oncogene* **2005**, *24*, 7410–7425. [CrossRef]

37. Zhao, Y.; Liu, Y.-S. Longevity Factor FOXO3: A Key Regulator in Aging-Related Vascular Diseases. *Front. Cardiovasc. Med.* **2021**, *8*, 778674. [CrossRef]

38. Kaletsky, R.; Lakhina, V.; Arey, R.; Williams, A.; Landis, J.; Ashraf, J.; Murphy, C.T. The *C. elegans* adult neuronal IIS/FOXO transcriptome reveals adult phenotype regulators. *Nature* **2016**, *529*, 92–96. [CrossRef]

39. Slack, C.; Giannakou, M.E.; Foley, A.; Goss, M.; Partridge, L. dFOXO-independent effects of reduced insulin-like signaling in Drosophila. *Aging Cell* **2011**, *10*, 735–748. [CrossRef]

40. Brenmoehl, J.; Hoeflich, A. Dual control of mitochondrial biogenesis by sirtuin 1 and sirtuin 3. *Mitochondrion* **2013**, *13*, 755–761. [CrossRef]

41. Chan, M.C.; Arany, Z. The Many roles of PGC-1α in Muscle—Recent Developments. *Metabolism* **2014**, *63*, 441–451. [CrossRef] [PubMed]

42. Summermatter, S.; Shui, G.; Maag, D.; Santos, G.; Wenk, M.R.; Handschin, C. PGC-1α improves glucose homeostasis in skeletal muscle in an activity-dependent manner. *Diabetes* **2013**, *62*, 85–95. [CrossRef] [PubMed]

43. Cantó, C.; Gerhart-Hines, Z.; Feige, J.N.; Lagouge, M.; Noriega, L.; Milne, J.C.; Elliott, P.J.; Puigserver, P.; Auwerx, J. AMPK regulates energy expenditure by modulating NAD$^+$ metabolism and SIRT1 activity. *Nature* **2009**, *458*, 1056–1060. [CrossRef] [PubMed]

44. Kauppinen, A.; Suuronen, T.; Ojala, J.; Kaarniranta, K.; Salminen, A. Antagonistic crosstalk between NF-κB and SIRT1 in the regulation of inflammation and metabolic disorders. *Cell. Signal.* **2013**, *25*, 1939–1948. [CrossRef] [PubMed]

45. Mao, K.; Zhang, G. The role of PARP1 in neurodegenerative diseases and aging. *FEBS J.* **2022**, *289*, 2013–2024. [CrossRef]

46. Amjad, S.; Nisar, S.; Bhat, A.A.; Shah, A.R.; Frenneaux, M.P.; Fakhro, K.; Haris, M.; Reddy, R.; Patay, Z.; Baur, J.; et al. Role of NAD+ in regulating cellular and metabolic signaling pathways. *Mol. Metab.* **2021**, *49*, 101195. [CrossRef]

47. Sedlackova, L.; Korolchuk, V.I. The crosstalk of NAD, ROS and autophagy in cellular health and ageing. *Biogerontology* **2020**, *21*, 381–397. [CrossRef]

48. Levine, D.C.; Hong, H.; Weidemann, B.J.; Ramsey, K.M.; Affinati, A.H.; Schmidt, M.S.; Cedernaes, J.; Omura, C.; Braun, R.; Lee, C.; et al. NAD$^+$ Controls Circadian Reprogramming through PER2 Nuclear Translocation to Counter Aging. *Mol. Cell* **2020**, *78*, 835–849.e7. [CrossRef]

49. Peek, C.B.; Affinati, A.H.; Ramsey, K.M.; Kuo, H.-Y.; Yu, W.; Sena, L.A.; Ilkayeva, O.; Marcheva, B.; Kobayashi, Y.; Omura, C.; et al. Circadian Clock NAD+ Cycle Drives Mitochondrial Oxidative Metabolism in Mice. *Science* **2013**, *342*, 1243417. [CrossRef]

50. Rehan, L.; Laszki-Szcząchor, K.; Sobieszczańska, M.; Polak-Jonkisz, D. SIRT1 and NAD as regulators of ageing. *Life Sci.* **2014**, *105*, 1–6. [CrossRef]

51. Chu, X.; Raju, R.P. Regulation of NAD$^+$ metabolism in aging and disease. *Metabolism* **2022**, *126*, 154923. [CrossRef] [PubMed]

52. Gilmour, B.C.; Gudmundsrud, R.; Frank, J.; Hov, A.; Lautrup, S.; Aman, Y.; Røsjø, H.; Brenner, C.; Ziegler, M.; Tysnes, O.-B.; et al. Targeting NAD+ in translational research to relieve diseases and conditions of metabolic stress and ageing. *Mech. Ageing Dev.* **2020**, *186*, 111208. [CrossRef] [PubMed]

53. Zhang, M.; Ying, W. NAD+ Deficiency Is a Common Central Pathological Factor of a Number of Diseases and Aging: Mechanisms and Therapeutic Implications. *Antioxid. Redox Signal.* **2019**, *30*, 890–905. [CrossRef]

54. Zhou, F.-Q. NAD$^+$, Senolytics, or Pyruvate for Healthy Aging? *Nutr. Metab. Insights* **2021**, *14*, 11786388211053408. [CrossRef] [PubMed]

55. Li, W.; Sauve, A.A. NAD$^+$ content and its role in mitochondria. *Methods Mol. Biol.* **2015**, *1241*, 39–48. [CrossRef]

56. Cerutti, R.; Pirinen, E.; Lamperti, C.; Marchet, S.; Sauve, A.A.; Li, W.; Leoni, V.; Schon, E.A.; Dantzer, F.; Auwerx, J.; et al. NAD(+)-dependent activation of Sirt1 corrects the phenotype in a mouse model of mitochondrial disease. *Cell Metab.* **2014**, *19*, 1042–1049. [CrossRef]

57. Li, W.; Li, Y.; Jin, X.; Liao, Q.; Chen, Z.; Peng, H.; Zhou, Y. CD38: A Significant Regulator of Macrophage Function. *Front. Oncol.* **2022**, *12*, 775649. [CrossRef]

58. Aksoy, P.; Escande, C.; White, T.A.; Thompson, M.; Soares, S.; Benech, J.C.; Chini, E.N. Regulation of SIRT 1 mediated NAD dependent deacetylation: A novel role for the multifunctional enzyme CD38. *Biochem. Biophys. Res. Commun.* **2006**, *349*, 353–359. [CrossRef]

59. Bu, X.; Kato, J.; Hong, J.A.; Merino, M.J.; Schrump, D.S.; Lund, F.E.; Moss, J. CD38 knockout suppresses tumorigenesis in mice and clonogenic growth of human lung cancer cells. *Carcinogenesis* **2018**, *39*, 242–251. [CrossRef]

60. Camacho-Pereira, J.; Tarragó, M.G.; Chini, C.C.S.; Nin, V.; Escande, C.; Warner, G.M.; Puranik, A.S.; Schoon, R.A.; Reid, J.M.; Galina, A.; et al. CD38 Dictates Age-Related NAD Decline and Mitochondrial Dysfunction through an SIRT3-Dependent Mechanism. *Cell Metab.* **2016**, *23*, 1127–1139. [CrossRef]

61. Domínguez-Pantoja, M.; López-Herrera, G.; Romero-Ramírez, H.; Santos-Argumedo, L.; Chávez-Rueda, A.K.; Hernández-Cueto, Á.; Flores-Muñoz, M.; Rodríguez-Alba, J.C. CD38 protein deficiency induces autoimmune characteristics and its activation enhances IL-10 production by regulatory B cells. *Scand. J. Immunol.* **2018**, *87*, e12664. [CrossRef]

62. Chini, C.C.S.; Peclat, T.R.; Warner, G.M.; Kashyap, S.; Espindola-Netto, J.M.; de Oliveira, G.C.; Gomez, L.S.; Hogan, K.A.; Tarragó, M.G.; Puranik, A.S.; et al. CD38 ecto-enzyme in immune cells is induced during aging and regulates NAD$^+$ and NMN levels. *Nat. Metab.* **2020**, *2*, 1284–1304. [CrossRef]

63. Chini, C.; Hogan, K.A.; Warner, G.M.; Tarragó, M.G.; Peclat, T.R.; Tchkonia, T.; Kirkland, J.L.; Chini, E. The NADase CD38 is induced by factors secreted from senescent cells providing a potential link between senescence and age-related cellular NAD+ decline. *Biochem. Biophys. Res. Commun.* **2019**, *513*, 486–493. [CrossRef]

64. Covarrubias, A.J.; Kale, A.; Perrone, R.; Lopez-Dominguez, J.A.; Pisco, A.O.; Kasler, H.G.; Schmidt, M.S.; Heckenbach, I.; Kwok, R.; Wiley, C.D.; et al. Senescent cells promote tissue NAD$^+$ decline during ageing via the activation of CD38$^+$ macrophages. *Nat. Metab.* **2020**, *2*, 1265–1283. [CrossRef]

65. Covarrubias, A.J.; Kale, A.; Perrone, R.; Lopez-Dominguez, J.A.; Pisco, A.O.; Kasler, H.G.; Schmidt, M.S.; Wiley, C.D.; Iyer, S.S.; Basisty, N.; et al. Aging-related inflammation driven by cellular senescence enhances NAD consumption via activation of CD38$^+$ pro-inflammatory macrophages. *bioRxiv* **2019**, 609438. [CrossRef]

66. Yoshino, J.; Baur, J.A.; Imai, S.-I. NAD$^+$ Intermediates: The Biology and Therapeutic Potential of NMN and NR. *Cell Metab.* **2018**, *27*, 513–528. [CrossRef]

67. Rajman, L.; Chwalek, K.; Sinclair, D.A. Therapeutic potential of NAD-boosting molecules: The in vivo evidence. *Cell Metab.* **2018**, *27*, 529–547. [CrossRef]

68. Roberts, M.D.; Osburn, S.C.; Godwin, J.S.; Ruple, B.A.; La Monica, M.B.; Raub, B.; Sandrock, J.E.; Ziegenfuss, T.N.; Lopez, H.L. Enhance Trial: Effects of NAD3® on Hallmarks of Aging and Clinical Endpoints of Health in Middle Aged Adults: A Subset Analysis Focused on Blood Cell NAD⁺ Concentrations and Lipid Metabolism. *Physiologia* **2022**, *2*, 20–31. [CrossRef]

69. Zhong, O.; Wang, J.; Tan, Y.; Lei, X.; Tang, Z. Effects of NAD+ precursor supplementation on glucose and lipid metabolism in humans: A meta-analysis. *Nutr. Metab.* **2022**, *19*, 20. [CrossRef]

70. Nadeeshani, H.; Li, J.; Ying, T.; Zhang, B.; Lu, J. Nicotinamide mononucleotide (NMN) as an anti-aging health product—Promises and safety concerns. *J. Adv. Res.* **2022**, *37*, 267–278. [CrossRef]

71. Shade, C. The Science Behind NMN—A Stable, Reliable NAD+Activator and Anti-Aging Molecule. *Integr. Med.* **2020**, *19*, 12–14.

72. Aman, Y.; Qiu, Y.; Tao, J.; Fang, E.F. Therapeutic potential of boosting NAD+ in aging and age-related diseases. *Transl. Med. Aging* **2018**, *2*, 30–37. [CrossRef]

73. Huang, H. A Multicentre, Randomised, Double Blind, Parallel Design, Placebo Controlled Study to Evaluate the Efficacy and Safety of Uthever (NMN Supplement), an Orally Administered Supplementation in Middle Aged and Older Adults. *Front. Aging* **2022**, *3*, 851698. [CrossRef] [PubMed]

74. Niu, K.-M.; Bao, T.; Gao, L.; Ru, M.; Li, Y.; Jiang, L.; Ye, C.; Wang, S.; Wu, X. The Impacts of Short-Term NMN Supplementation on Serum Metabolism, Fecal Microbiota, and Telomere Length in Pre-Aging Phase. *Front. Nutr.* **2021**, *8*, 756243. [CrossRef] [PubMed]

75. Huang, P.; Jiang, A.; Wang, X.; Zhou, Y.; Tang, W.; Ren, C.; Qian, X.; Zhou, Z.; Gong, A. NMN Maintains Intestinal Homeostasis by Regulating the Gut Microbiota. *Front. Nutr.* **2021**, *8*, 714604. [CrossRef] [PubMed]

76. Kiss, T.; Giles, C.B.; Tarantini, S.; Yabluchanskiy, A.; Balasubramanian, P.; Gautam, T.; Csipo, T.; Nyúl-Tóth, Á.; Lipecz, A.; Szabo, C.; et al. Nicotinamide mononucleotide (NMN) supplementation promotes anti-aging miRNA expression profile in the aorta of aged mice, predicting epigenetic rejuvenation and anti-atherogenic effects. *Geroscience* **2019**, *41*, 419–439. [CrossRef] [PubMed]

77. Liao, B.; Zhao, Y.; Wang, D.; Zhang, X.; Hao, X.; Hu, M. Nicotinamide mononucleotide supplementation enhances aerobic capacity in amateur runners: A randomized, double-blind study. *J. Int. Soc. Sport. Nutr.* **2021**, *18*, 54. [CrossRef]

78. Igarashi, M.; Nakagawa-Nagahama, Y.; Miura, M.; Kashiwabara, K.; Yaku, K.; Sawada, M.; Sekine, R.; Fukamizu, Y.; Sato, T.; Sakurai, T.; et al. Chronic nicotinamide mononucleotide supplementation elevates blood nicotinamide adenine dinucleotide levels and alters muscle function in healthy older men. *npj Aging* **2022**, *8*, 5. [CrossRef]

79. Kim, M.; Seol, J.; Sato, T.; Fukamizu, Y.; Sakurai, T.; Okura, T. Effect of 12-Week Intake of Nicotinamide Mononucleotide on Sleep Quality, Fatigue, and Physical Performance in Older Japanese Adults: A Randomized, Double-Blind Placebo-Controlled Study. *Nutrients* **2022**, *14*, 755. [CrossRef]

80. Maric, T.; Bazhin, A.; Khodakivskyi, P.; Mikhaylov, G.; Solodnikova, E.; Yevtodiyenko, A.; Giordano Attianese, G.M.P.; Coukos, G.; Irving, M.; Joffraud, M.; et al. A bioluminescent-based probe for in vivo non-invasive monitoring of nicotinamide riboside uptake reveals a link between metastasis and NAD+ metabolism. *Biosens. Bioelectron.* **2023**, *220*, 114826. [CrossRef]

81. Cros, C.; Cannelle, H.; Laganier, L.; Grozio, A.; Canault, M. Safety evaluation after acute and sub-chronic oral administration of high purity nicotinamide mononucleotide (NMN-C®) in Sprague-Dawley rats. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* **2021**, *150*, 112060. [CrossRef]

82. Yoshino, M.; Yoshino, J.; Kayser, B.D.; Patti, G.J.; Franczyk, M.P.; Mills, K.F.; Sindelar, M.; Pietka, T.; Patterson, B.W.; Imai, S.-I.; et al. Nicotinamide mononucleotide increases muscle insulin sensitivity in prediabetic women. *Science* **2021**, *372*, 1224–1229. [CrossRef]

83. Irie, J.; Inagaki, E.; Fujita, M.; Nakaya, H.; Mitsuishi, M.; Yamaguchi, S.; Yamashita, K.; Shigaki, S.; Ono, T.; Yukioka, H.; et al. Effect of oral administration of nicotinamide mononucleotide on clinical parameters and nicotinamide metabolite levels in healthy Japanese men. *Endocr. J.* **2020**, *67*, 153–160. [CrossRef]

84. Fukamizu, Y.; Uchida, Y.; Shigekawa, A.; Sato, T.; Kosaka, H.; Sakurai, T. Safety evaluation of β-nicotinamide mononucleotide oral administration in healthy adult men and women. *Sci. Rep.* **2022**, *12*, 14442. [CrossRef]

85. Yi, L.; Maier, A.B.; Tao, R.; Lin, Z.; Vaidya, A.; Pendse, S.; Thasma, S.; Andhalkar, N.; Avhad, G.; Kumbhar, V. The efficacy and safety of β-nicotinamide mononucleotide (NMN) supplementation in healthy middle-aged adults: A randomized, multicenter, double-blind, placebo-controlled, parallel-group, dose-dependent clinical trial. *GeroScience* **2022**, 1–15. [CrossRef]

86. Pencina, K.; Lavu, S.; Dos Santos, M.; Beleva, Y.M.; Cheng, M.; Livingston, D.; Bhasin, S. MIB-626, an Oral Formulation of a Microcrystalline Unique Polymorph of β-Nicotinamide Mononucleotide, Increases Circulating Nicotinamide Adenine Dinucleotide and its Metabolome in Middle-aged and Older Adults. *J. Gerontol. A Biol. Sci. Med. Sci.* **2022**, glac049. [CrossRef]

87. Tomé-Carneiro, J.; Larrosa, M.; González-Sarrías, A.; Tomás-Barberán, F.A.; García-Conesa, M.T.; Espín, J.C. Resveratrol and Clinical Trials: The Crossroad from In Vitro Studies to Human Evidence. *Curr. Pharm. Des.* **2013**, *19*, 6064–6093. [CrossRef]

88. Resveratrol and pterostilbene: A comparative overview of their chemistry, biosynthesis, plant sources and pharmacological properties. *J. App. Pharm. Sci.* **2019**, *9*, 124–129. [CrossRef]

89. Riche, D.M.; McEwen, C.L.; Riche, K.D.; Sherman, J.J.; Wofford, M.R.; Deschamp, D.; Griswold, M. Analysis of Safety from a Human Clinical Trial with Pterostilbene. *J. Toxicol.* **2013**, *2013*, e463595. [CrossRef]

90. Ramírez-Garza, S.L.; Laveriano-Santos, E.P.; Marhuenda-Muñoz, M.; Storniolo, C.E.; Tresserra-Rimbau, A.; Vallverdú-Queralt, A.; Lamuela-Raventós, R.M. Health Effects of Resveratrol: Results from Human Intervention Trials. *Nutrients* **2018**, *10*, 1892. [CrossRef]

91. Li, Y.-R.; Li, S.; Lin, C.-C. Effect of resveratrol and pterostilbene on aging and longevity. *Biofactors* **2018**, *44*, 69–82. [CrossRef] [PubMed]

92. Beghelli, D.; Zallocco, L.; Barbalace, M.C.; Paglia, S.; Strocchi, S.; Cirilli, I.; Marzano, V.; Putignani, L.; Lupidi, G.; Hrelia, S.; et al. Pterostilbene Promotes Mean Lifespan in Both Male and Female Drosophila Melanogaster Modulating Different Proteins in the Two Sexes. *Oxid. Med. Cell. Longev.* **2022**, *2022*, 1744408. [CrossRef] [PubMed]

93. McCormack, D.; McFadden, D. A Review of Pterostilbene Antioxidant Activity and Disease Modification. *Oxid. Med. Cell. Longev.* **2013**, *2013*, 575482. [CrossRef] [PubMed]

94. Knutson, M.D.; Leeuwenburgh, C. Resveratrol and novel potent activators of SIRT1: Effects on aging and age-related diseases. *Nutr. Rev.* **2008**, *66*, 591–596. [CrossRef] [PubMed]

95. Yabluchanskiy, A.; Ma, Y.; Iyer, R.P.; Hall, M.E.; Lindsey, M.L. Matrix Metalloproteinase-9: Many Shades of Function in Cardiovascular Disease. *Physiology* **2013**, *28*, 391–403. [CrossRef]

96. Moens, U.; Kostenko, S.; Sveinbjørnsson, B. The Role of Mitogen-Activated Protein Kinase-Activated Protein Kinases (MAP-KAPKs) in Inflammation. *Genes* **2013**, *4*, 101–133. [CrossRef]

97. De la Lastra, C.A.; Villegas, I. Resveratrol as an anti-inflammatory and anti-aging agent: Mechanisms and clinical implications. *Mol. Nutr. Food Res.* **2005**, *49*, 405–430. [CrossRef]

98. Teng, W.-L.; Huang, P.-H.; Wang, H.-C.; Tseng, C.-H.; Yen, F.-L. Pterostilbene Attenuates Particulate Matter-Induced Oxidative Stress, Inflammation and Aging in Keratinocytes. *Antioxidants* **2021**, *10*, 1552. [CrossRef]

99. Kasiotis, K.M.; Pratsinis, H.; Kletsas, D.; Haroutounian, S.A. Resveratrol and related stilbenes: Their anti-aging and anti-angiogenic properties. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* **2013**, *61*, 112–120. [CrossRef]

100. Hecker, A.; Schellnegger, M.; Hofmann, E.; Luze, H.; Nischwitz, S.P.; Kamolz, L.; Kotzbeck, P. The impact of resveratrol on skin wound healing, scarring, and aging. *Int. Wound J.* **2021**, *19*, 9–28. [CrossRef]

101. Escolme, S.M.; Wakeling, L.A.; Alatawi, F.; Valentine, R.; Ford, D. Does resveratrol act independently of SIRT1 to affect genes relevant to ageing? *Proc. Nutr. Soc.* **2013**, *72*, E191. [CrossRef]

102. Fernández, A.F.; Fraga, M.F. The effects of the dietary polyphenol resveratrol on human healthy aging and lifespan. *Epigenetics* **2011**, *6*, 870–874. [CrossRef]

103. Hu, Y.; Liu, J.; Wang, J.; Liu, Q. The controversial links among calorie restriction, SIRT1, and resveratrol. *Free Radic. Biol. Med.* **2011**, *51*, 250–256. [CrossRef]

104. Bonkowski, M.S.; Sinclair, D.A. Slowing ageing by design: The rise of NAD+ and sirtuin-activating compounds. *Nat. Rev. Mol. Cell Biol.* **2016**, *17*, 679–690. [CrossRef]

105. Zhou, D.-D.; Luo, M.; Huang, S.-Y.; Saimaiti, A.; Shang, A.; Gan, R.-Y.; Li, H.-B. Effects and Mechanisms of Resveratrol on Aging and Age-Related Diseases. *Oxid. Med. Cell. Longev.* **2021**, *2021*, 9932218. [CrossRef]

106. Lachiondo-Ortega, S.; Delgado, T.C.; Baños-Jaime, B.; Velázquez-Cruz, A.; Díaz-Moreno, I.; Martínez-Chantar, M.L. Hu Antigen R (HuR) Protein Structure, Function and Regulation in Hepatobiliary Tumors. *Cancers* **2022**, *14*, 2666. [CrossRef]

107. Barger, J.L.; Kayo, T.; Vann, J.M.; Arias, E.B.; Wang, J.; Hacker, T.A.; Wang, Y.; Raederstorff, D.; Morrow, J.D.; Leeuwenburgh, C.; et al. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS ONE* **2008**, *3*, e2264. [CrossRef]

108. Li, J.; Zhang, C.-X.; Liu, Y.-M.; Chen, K.-L.; Chen, G. A comparative study of anti-aging properties and mechanism: Resveratrol and caloric restriction. *Oncotarget* **2017**, *8*, 65717–65729. [CrossRef]

109. Uddin, M.J.; Farjana, M.; Moni, A.; Hossain, K.S.; Hannan, M.A.; Ha, H. Prospective Pharmacological Potential of Resveratrol in Delaying Kidney Aging. *Int. J. Mol. Sci.* **2021**, *22*, 8258. [CrossRef]

110. Kim, E.N.; Kim, M.Y.; Lim, J.H.; Kim, Y.; Shin, S.J.; Park, C.W.; Kim, Y.-S.; Chang, Y.S.; Yoon, H.E.; Choi, B.S. The protective effect of resveratrol on vascular aging by modulation of the renin-angiotensin system. *Atherosclerosis* **2018**, *270*, 123–131. [CrossRef]

111. Caldeira, C.A.; Santos, M.A.; Araújo, G.R.; Lara, R.C.; Franco, F.N.; Chaves, M.M. Resveratrol: Change of SIRT 1 and AMPK signaling pattern during the aging process. *Exp. Gerontol.* **2021**, *146*, 111226. [CrossRef] [PubMed]

112. Demidenko, Z.N.; Blagosklonny, M.V. At concentrations that inhibit mTOR, resveratrol suppresses cellular senescence. *Cell Cycle* **2009**, *8*, 1901–1904. [CrossRef] [PubMed]

113. Zhou, J.; Xue, Z.; He, H.-N.; Liu, X.; Yin, S.-Y.; Wu, D.-Y.; Zhang, X.; Schatten, H.; Miao, Y.-L. Resveratrol delays postovulatory aging of mouse oocytes through activating mitophagy. *Aging* **2019**, *11*, 11504–11519. [CrossRef]

114. Sun, Y.-L.; Tang, S.-B.; Shen, W.; Yin, S.; Sun, Q.-Y. Roles of Resveratrol in Improving the Quality of Postovulatory Aging Oocytes In Vitro. *Cells* **2019**, *8*, 1132. [CrossRef] [PubMed]

115. Chandrasekaran, K.; Salimian, M.; Konduru, S.R.; Choi, J.; Kumar, P.; Long, A.; Klimova, N.; Ho, C.-Y.; Kristian, T.; Russell, J.W. Overexpression of Sirtuin 1 protein in neurons prevents and reverses experimental diabetic neuropathy. *Brain* **2019**, *142*, 3737–3752. [CrossRef]

116. Moraes, D.S.; Moreira, D.C.; Andrade, J.M.O.; Santos, S.H.S. Sirtuins, brain and cognition: A review of resveratrol effects. *IBRO Rep.* **2020**, *9*, 46–51. [CrossRef]

117. Gocmez, S.S.; Gacar, N.; Utkan, T.; Gacar, G.; Scarpace, P.J.; Tumer, N. Protective effects of resveratrol on aging-induced cognitive impairment in rats. *Neurobiol. Learn Mem.* **2016**, *131*, 131–136. [CrossRef]

118. Oomen, C.A.; Farkas, E.; Roman, V.; Van Der Beek, E.M.; Luiten, P.G.; Meerlo, P. Resveratrol preserves cerebrovascular density and cognitive function in aging mice. *Front. Aging Neurosci.* **2009**, *1*, 4. [CrossRef]

119. Szczurek-Janicka, P.; Ropka-Molik, K.; Oczkowicz, M.; Orczewska-Dudek, S.; Pietras, M.; Pieszka, M. Expression Profile of Brain Aging and Metabolic Function are Altered by Resveratrol or α-Ketoglutarate Supplementation in Rats Fed a High-Fat Diet. *Pol. J. Food Nutr. Sci.* **2021**, *71*, 255–268. [CrossRef]

120. Dubowitz, N.; Xue, W.; Long, Q.; Ownby, J.G.; Olson, D.E.; Barb, D.; Rhee, M.K.; Mohan, A.V.; Watson-Williams, P.I.; Jackson, S.L.; et al. Aging is associated with increased HbA1c levels, independently of glucose levels and insulin resistance, and also with decreased HbA1c diagnostic specificity. *Diabet. Med. J. Br. Diabet. Assoc.* **2014**, *31*, 927–935. [CrossRef]

121. Witte, A.V.; Kerti, L.; Margulies, D.S.; Flöel, A. Effects of Resveratrol on Memory Performance, Hippocampal Functional Connectivity, and Glucose Metabolism in Healthy Older Adults. *J. Neurosci.* **2014**, *34*, 7862–7870. [CrossRef]

122. Moussa, C.; Hebron, M.; Huang, X.; Ahn, J.; Rissman, R.A.; Aisen, P.S.; Turner, R.S. Resveratrol regulates neuro-inflammation and induces adaptive immunity in Alzheimer's disease. *J. Neuroinflam.* **2017**, *14*, 1. [CrossRef]

123. Mahjabeen, W.; Khan, D.A.; Mirza, S.A. Role of resveratrol supplementation in regulation of glucose hemostasis, inflammation and oxidative stress in patients with diabetes mellitus type 2: A randomized, placebo-controlled trial. *Complement. Ther. Med.* **2022**, *66*, 102819. [CrossRef]

124. Wong, R.H.; Thaung Zaw, J.J.; Xian, C.J.; Howe, P.R. Regular Supplementation with Resveratrol Improves Bone Mineral Density in Postmenopausal Women: A Randomized, Placebo-Controlled Trial. *J. Bone Miner. Res. Off. J. Am. Soc. Bone Miner. Res.* **2020**, *35*, 2121–2131. [CrossRef]

125. Thaung Zaw, J.J.; Howe, P.R.C.; Wong, R.H.X. Long-term effects of resveratrol on cognition, cerebrovascular function and cardio-metabolic markers in postmenopausal women: A 24-month randomised, double-blind, placebo-controlled, crossover study. *Clin. Nutr.* **2021**, *40*, 820–829. [CrossRef]

126. Zaw, J.J.T.; Howe, P.R.C.; Wong, R.H.X. Sustained Cerebrovascular and Cognitive Benefits of Resveratrol in Postmenopausal Women. *Nutrients* **2020**, *12*, 828. [CrossRef]

127. Mo, Z.; Zheng, S.; Lv, Z.; Zhuang, Y.; Lan, X.; Wang, F.; Lu, X.; Zhao, Y.; Zhou, S. Senescence marker protein 30 (SMP30) serves as a potential prognostic indicator in hepatocellular carcinoma. *Sci. Rep.* **2016**, *6*, 39376. [CrossRef]

128. Torregrosa-Muñumer, R.; Vara, E.; Fernández-Tresguerres, J.Á.; Gredilla, R. Resveratrol supplementation at old age reverts changes associated with aging in inflammatory, oxidative and apoptotic markers in rat heart. *Eur. J. Nutr.* **2021**, *60*, 2683–2693. [CrossRef]

129. Smoliga, J.M.; Baur, J.; Hausenblas, H.A. Resveratrol and health—A comprehensive review of human clinical trials. *Mol. Nutr. Food Res.* **2011**, *55*, 1129–1141. [CrossRef]

130. Mehta, S.; Young, C.C.; Warren, M.R.; Akhtar, S.; Shefelbine, S.J.; Crane, J.D.; Bajpayee, A.G. Resveratrol and Curcumin Attenuate Ex Vivo Sugar-Induced Cartilage Glycation, Stiffening, Senescence, and Degeneration. *Cartilage* **2021**, *13*, 1214S–1228S. [CrossRef]

131. Furuyama, T.; Kitayama, K.; Yamashita, H.; Mori, N. Forkhead transcription factor FOXO1 (FKHR)-dependent induction of PDK4 gene expression in skeletal muscle during energy deprivation. *Biochem. J.* **2003**, *375*, 365–371. [CrossRef] [PubMed]

132. Sin, T.K.; Yung, B.Y.; Siu, P.M. Modulation of SIRT1-Foxo1 signaling axis by resveratrol: Implications in skeletal muscle aging and insulin resistance. *Cell Physiol. Biochem. Int. J. Exp. Cell Physiol. Biochem. Pharmacol.* **2015**, *35*, 541–552. [CrossRef] [PubMed]

133. Ramis, M.R.; Esteban, S.; Miralles, A.; Tan, D.-X.; Reiter, R.J. Caloric restriction, resveratrol and melatonin: Role of SIRT1 and implications for aging and related-diseases. *Mech. Ageing Dev.* **2015**, *146–148*, 28–41. [CrossRef] [PubMed]

134. Corrêa, R.C.G.; Peralta, R.M.; Haminiuk, C.W.I.; Maciel, G.M.; Bracht, A.; Ferreira, I.C.F.R. New phytochemicals as potential human anti-aging compounds: Reality, promise, and challenges. *Crit. Rev. Food Sci. Nutr.* **2018**, *58*, 942–957. [CrossRef] [PubMed]

135. Smoliga, J.M.; Colombo, E.S.; Campen, M.J. A healthier approach to clinical trials evaluating resveratrol for primary prevention of age-related diseases in healthy populations. *Aging* **2013**, *5*, 495–506. [CrossRef] [PubMed]

136. Bai, L.; Yau, L.; Tong, T.; Chan, W.; Zhang, W.; Jiang, Z. Improvement of tissue-specific distribution and biotransformation potential of nicotinamide mononucleotide in combination with ginsenosides or resveratrol. *Pharmacol. Res. Perspect.* **2022**, *10*, e00986. [CrossRef] [PubMed]

137. Grant, R. Resveratrol Increases Intracellular NAD$^+$ Levels Through Up regulation of the NAD$^+$ Synthetic Enzyme Nicotinamide Mononucleotide Adenylyltransferase. *Nat. Preced.* **2010**, *4421*, 1–8. [CrossRef]

138. Simic, P.; Vela Parada, X.F.; Parikh, S.M.; Dellinger, R.; Guarente, L.P.; Rhee, E.P. Nicotinamide riboside with pterostilbene (NRPT) increases NAD$^+$ in patients with acute kidney injury (AKI): A randomized, double-blind, placebo-controlled, stepwise safety study of escalating doses of NRPT in patients with AKI. *BMC Nephrol.* **2020**, *21*, 342. [CrossRef]

139. Dellinger, R.W.; Santos, S.R.; Morris, M.; Evans, M.; Alminana, D.; Guarente, L.; Marcotulli, E. Repeat dose NRPT (nicotinamide riboside and pterostilbene) increases NAD$^+$ levels in humans safely and sustainably: A randomized, double-blind, placebo-controlled study. *npj Aging Mech. Dis.* **2017**, *3*, 17. [CrossRef]

140. Yang, Y.; Liu, Z.; Zhao, B.; Joseph, S.; Ma, Y.; Xu, H.; Cheng, H. Resveratrol and Quercetin Potentiate the Cell Protection and Rescue Effects of NAD+ Precursors in HEK293 Cells Challenged by DNA Damaging Agent, N-Methyl-N′-nitro-N-nitrosoguanidine. *Nat. Prod. Commun.* **2021**, *16*, 1934578X211045465. [CrossRef]

141. Garrido-Maraver, J.; Cordero, M.D.; Oropesa-Ávila, M.; Fernández Vega, A.; de la Mata, M.; Delgado Pavón, A.; de Miguel, M.; Pérez Calero, C.; Villanueva Paz, M.; Cotán, D.; et al. Coenzyme q10 therapy. *Mol. Syndromol.* **2014**, *5*, 187–197. [CrossRef]

142. Garrido-Maraver, J.; Cordero, M.D.; Oropesa-Avila, M.; Vega, A.F.; de la Mata, M.; Pavon, A.D.; Alcocer-Gomez, E.; Calero, C.P.; Paz, M.V.; Alanis, M.; et al. Clinical applications of coenzyme Q$_{10}$. *Front. Biosci.* **2014**, *19*, 619–633. [CrossRef]

143. Martelli, A.; Testai, L.; Colletti, A.; Cicero, A.F.G. Coenzyme Q10: Clinical Applications in Cardiovascular Diseases. *Antioxidants* **2020**, *9*, 341. [CrossRef]

144. Hernández-Camacho, J.D.; Bernier, M.; López-Lluch, G.; Navas, P. Coenzyme Q10 Supplementation in Aging and Disease. *Front. Physiol.* **2018**, *9*, 44. [CrossRef]

145. Gutierrez-Mariscal, F.M.; Yubero-Serrano, E.M.; Villalba, J.M.; Lopez-Miranda, J. Coenzyme Q10: From bench to clinic in aging diseases, a translational review. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 2240–2257. [CrossRef] [PubMed]

146. Díaz-Casado, M.E.; Quiles, J.L.; Barriocanal-Casado, E.; González-García, P.; Battino, M.; López, L.C.; Varela-López, A. The Paradox of Coenzyme Q10 in Aging. *Nutrients* **2019**, *11*, 2221. [CrossRef]

147. González-Guardia, L.; Yubero-Serrano, E.M.; Delgado-Lista, J.; Perez-Martinez, P.; Garcia-Rios, A.; Marin, C.; Camargo, A.; Delgado-Casado, N.; Roche, H.M.; Perez-Jimenez, F.; et al. Effects of the Mediterranean diet supplemented with coenzyme q10 on metabolomic profiles in elderly men and women. *J. Gerontol. A Biol. Sci. Med. Sci.* **2015**, *70*, 78–84. [CrossRef] [PubMed]

148. Castro-Marrero, J.; Sáez-Francàs, N.; Segundo, M.J.; Calvo, N.; Faro, M.; Aliste, L.; Fernández de Sevilla, T.; Alegre, J. Effect of coenzyme Q10 plus nicotinamide adenine dinucleotide supplementation on maximum heart rate after exercise testing in chronic fatigue syndrome—A randomized, controlled, double-blind trial. *Clin. Nutr.* **2016**, *35*, 826–834. [CrossRef]

149. Castro-Marrero, J.; Cordero, M.D.; Segundo, M.J.; Sáez-Francàs, N.; Calvo, N.; Román-Malo, L.; Aliste, L.; Fernández de Sevilla, T.; Alegre, J. Does oral coenzyme Q10 plus NADH supplementation improve fatigue and biochemical parameters in chronic fatigue syndrome? *Antioxid. Redox Signal.* **2015**, *22*, 679–685. [CrossRef]

150. Zeisel, S. Choline, Other Methyl-Donors and Epigenetics. *Nutrients* **2017**, *9*, 445. [CrossRef]

151. Go, E.K.; Jung, K.J.; Kim, J.Y.; Yu, B.P.; Chung, H.Y. Betaine suppresses proinflammatory signaling during aging: The involvement of nuclear factor-kappaB via nuclear factor-inducing kinase/IkappaB kinase and mitogen-activated protein kinases. *J. Gerontol. A Biol. Sci. Med. Sci.* **2005**, *60*, 1252–1264. [CrossRef]

152. Zhao, G.; He, F.; Wu, C.; Li, P.; Li, N.; Deng, J.; Zhu, G.; Ren, W.; Peng, Y. Betaine in Inflammation: Mechanistic Aspects and Applications. *Front. Immunol.* **2018**, *9*, 1070. [CrossRef] [PubMed]

153. Sun, W.-P.; Zhai, M.-Z.; Li, D.; Zhou, Y.; Chen, N.-N.; Guo, M.; Zhou, S.-S. Comparison of the effects of nicotinic acid and nicotinamide degradation on plasma betaine and choline levels. *Clin. Nutr.* **2017**, *36*, 1136–1142. [CrossRef] [PubMed]

154. Li, D.; Tian, Y.-J.; Guo, J.; Sun, W.-P.; Lun, Y.-Z.; Guo, M.; Luo, N.; Cao, Y.; Cao, J.-M.; Gong, X.-J.; et al. Nicotinamide supplementation induces detrimental metabolic and epigenetic changes in developing rats. *Br. J. Nutr.* **2013**, *110*, 2156–2164. [CrossRef] [PubMed]

155. Kashyap, D.; Garg, V.K.; Tuli, H.S.; Yerer, M.B.; Sak, K.; Sharma, A.K.; Kumar, M.; Aggarwal, V.; Sandhu, S.S. Fisetin and Quercetin: Promising Flavonoids with Chemopreventive Potential. *Biomolecules* **2019**, *9*, 174. [CrossRef]

156. Yousefzadeh, M.J.; Zhu, Y.; McGowan, S.J.; Angelini, L.; Fuhrmann-Stroissnigg, H.; Xu, M.; Ling, Y.Y.; Melos, K.I.; Pirtskhalava, T.; Inman, C.L.; et al. Fisetin is a senotherapeutic that extends health and lifespan. *eBioMedicine* **2018**, *36*, 18–28. [CrossRef]

157. Jang, S.-Y.; Kang, H.T.; Hwang, E.S. Nicotinamide-induced Mitophagy. *J. Biol. Chem.* **2012**, *287*, 19304–19314. [CrossRef]

158. Deepika; Maurya, P.K. Health Benefits of Quercetin in Age-Related Diseases. *Molecules* **2022**, *27*, 2498. [CrossRef]

159. Geng, L.; Liu, Z.; Zhang, W.; Li, W.; Wu, Z.; Wang, W.; Ren, R.; Su, Y.; Wang, P.; Sun, L.; et al. Chemical screen identifies a geroprotective role of quercetin in premature aging. *Protein Cell* **2018**, *10*, 417–435. [CrossRef]

160. Escande, C.; Nin, V.; Price, N.L.; Capellini, V.; Gomes, A.P.; Barbosa, M.T.; O'Neil, L.; White, T.A.; Sinclair, D.A.; Chini, E.N. Flavonoid Apigenin Is an Inhibitor of the NAD+ase CD38. *Diabetes* **2013**, *62*, 1084–1093. [CrossRef]

161. Zhang, F.; Feng, J.; Zhang, J.; Kang, X.; Qian, D. Quercetin modulates AMPK/SIRT1/NF-κB signaling to inhibit inflammatory/oxidative stress responses in diabetic high fat diet-induced atherosclerosis in the rat carotid artery. *Exp. Ther. Med.* **2020**, *20*, 280. [CrossRef]

162. Buss, G.D.; Constantin, J.; de Lima, L.C.N.; Teodoro, G.R.; Comar, J.F.; Ishii-Iwamoto, E.L.; Bracht, A. The Action of Quercetin on the Mitochondrial NADH to NAD$^+$ Ratio in the Isolated Perfused Rat Liver. *Planta Med.* **2005**, *71*, 1118–1122. [CrossRef]

163. Gendrisch, F.; Esser, P.R.; Schempp, C.M.; Wölfle, U. Luteolin as a modulator of skin aging and inflammation. *BioFactors* **2021**, *47*, 170–180. [CrossRef]

164. Compound in Celery, Peppers Reduces Age-Related Memory Deficits. Available online: https://www.sciencedaily.com/releases/2010/10/101013122601.htm (accessed on 11 November 2022).

165. Kellenberger, E.; Kuhn, I.; Schuber, F.; Muller-Steffner, H. Flavonoids as inhibitors of human CD38. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3939–3942. [CrossRef]

166. Boslett, J.; Hemann, C.; Zhao, Y.J.; Lee, H.-C.; Zweier, J.L. Luteolinidin Protects the Postischemic Heart through CD38 Inhibition with Preservation of NAD(P)(H). *J. Pharmacol. Exp. Ther.* **2017**, *361*, 99–108. [CrossRef]

167. Wissler Gerdes, E.O.; Misra, A.; Netto, J.M.E.; Tchkonia, T.; Kirkland, J.L. Strategies for late phase preclinical and early clinical trials of senolytics. *Mech. Ageing Dev.* **2021**, *200*, 111591. [CrossRef]

168. Farkas, O.; Palócz, O.; Pászti-Gere, E.; Gálfi, P. Polymethoxyflavone Apigenin-Trimethylether Suppresses LPS-Induced Inflammatory Response in Nontransformed Porcine Intestinal Cell Line IPEC-J2. *Oxidative Med. Cell. Longev.* **2015**, *2015*, e673847. [CrossRef]

169. Ali, F.; Rahul; Naz, F.; Jyoti, S.; Siddique, Y.H. Health functionality of apigenin: A review. *Int. J. Food Prop.* **2017**, *20*, 1197–1238. [CrossRef]

170. Ahmed, S.A.; Parama, D.; Daimari, E.; Girisa, S.; Banik, K.; Harsha, C.; Dutta, U.; Kunnumakkara, A.B. Rationalizing the therapeutic potential of apigenin against cancer. *Life Sci.* **2021**, *267*, 118814. [CrossRef]

171. Qiu, J.-G.; Wang, L.; Liu, W.-J.; Wang, J.-F.; Zhao, E.-J.; Zhou, F.-M.; Ji, X.-B.; Wang, L.-H.; Xia, Z.-K.; Wang, W.; et al. Apigenin Inhibits IL-6 Transcription and Suppresses Esophageal Carcinogenesis. *Front. Pharmacol.* **2019**, *10*, 1002. [CrossRef]

172. Budhraja, A.; Gao, N.; Zhang, Z.; Son, Y.-O.; Cheng, S.; Wang, X.; Ding, S.; Hitron, A.; Chen, G.; Luo, J.; et al. Apigenin induces apoptosis in human leukemia cells and exhibits anti-leukemic activity in vivo. *Mol. Cancer Ther.* **2012**, *11*, 132–142. [CrossRef] [PubMed]

173. Alam, W.; Rocca, C.; Khan, H.; Hussain, Y.; Aschner, M.; De Bartolo, A.; Amodio, N.; Angelone, T.; Cheang, W.S. Current Status and Future Perspectives on Therapeutic Potential of Apigenin: Focus on Metabolic-Syndrome-Dependent Organ Dysfunction. *Antioxidants* **2021**, *10*, 1643. [CrossRef] [PubMed]

174. Ginwala, R.; Bhavsar, R.; Chigbu, D.G.I.; Jain, P.; Khan, Z.K. Potential Role of Flavonoids in Treating Chronic Inflammatory Diseases with a Special Focus on the Anti-Inflammatory Activity of Apigenin. *Antioxidants* **2019**, *8*, 35. [CrossRef] [PubMed]

175. Choi, W.H.; Son, H.J.; Jang, Y.J.; Ahn, J.; Jung, C.H.; Ha, T.Y. Apigenin Ameliorates the Obesity-Induced Skeletal Muscle Atrophy by Attenuating Mitochondrial Dysfunction in the Muscle of Obese Mice. *Mol. Nutr. Food Res.* **2017**, *61*, 1700218. [CrossRef]

176. Su, T.; Chunhua, H.; Yang, C.; Jiang, T.; Su, J.; Chen, M.; Fatima, S.; Gong, R.-H.; Hu, X.; Bian, Z.-X.; et al. Apigenin inhibits STAT3/CD36 signaling axis and reduces visceral obesity. *Pharmacol. Res.* **2019**, *152*, 104586. [CrossRef]

177. Clayton, Z.S.; Hutton, D.A.; Brunt, V.E.; VanDongen, N.S.; Ziemba, B.P.; Casso, A.G.; Greenberg, N.T.; Mercer, A.N.; Rossman, M.J.; Campisi, J.; et al. Apigenin restores endothelial function by ameliorating oxidative stress, reverses aortic stiffening, and mitigates vascular inflammation with aging. *Am. J. Physiol. Heart Circ. Physiol.* **2021**, *321*, H185–H196. [CrossRef]

178. Sharma, A.; Ghani, A.; Sak, K.; Tuli, H.S.; Sharma, A.K.; Setzer, W.N.; Sharma, S.; Das, A.K. Probing into Therapeutic Anti-cancer Potential of Apigenin: Recent Trends and Future Directions. *Recent. Pat. Inflamm. Allergy Drug Discov.* **2019**, *13*, 124–133. [CrossRef]

179. Li, B.S.; Zhu, R.Z.; Lim, S.-H.; Seo, J.H.; Choi, B.-M. Apigenin Alleviates Oxidative Stress-Induced Cellular Senescence via Modulation of the SIRT1-NAD+-CD38 Axis. *Am. J. Chin. Med.* **2021**, *49*, 1235–1250. [CrossRef]

180. Ogura, Y.; Kitada, M.; Xu, J.; Monno, I.; Koya, D. CD38 inhibition by apigenin ameliorates mitochondrial oxidative stress through restoration of the intracellular NAD+/NADH ratio and Sirt3 activity in renal tubular cells in diabetic rats. *Aging* **2020**, *12*, 11325–11336. [CrossRef]

181. Sztretye, M.; Dienes, B.; Gönczi, M.; Czirják, T.; Csernoch, L.; Dux, L.; Szentesi, P.; Keller-Pintér, A. Astaxanthin: A Potential Mitochondrial-Targeted Antioxidant Treatment in Diseases and with Aging. *Oxid. Med. Cell. Longev.* **2019**, *2019*, 3849692. [CrossRef]

182. Zhang, X.-S.; Lu, Y.; Li, W.; Tao, T.; Peng, L.; Wang, W.-H.; Gao, S.; Liu, C.; Zhuang, Z.; Xia, D.-Y.; et al. Astaxanthin ameliorates oxidative stress and neuronal apoptosis via SIRT1/NRF2/Prx2/ASK1/p38 after traumatic brain injury in mice. *Br. J. Pharmacol.* **2021**, *178*, 1114–1132. [CrossRef]

183. Zhang, J.; Wang, Q.; Zhao, S.; Ji, X.; Qiu, J.; Wang, J.; Zhou, Y.; Cai, Q.; Zhang, J.; Gao, H. Astaxanthin attenuated pressure overload-induced cardiac dysfunction and myocardial fibrosis: Partially by activating SIRT1. *Biochim. Biophys. Acta BBA Gen. Subj.* **2017**, *1861*, 1715–1728. [CrossRef]

184. Shatoor, A.S.; Al Humayed, S. Astaxanthin Ameliorates high-fat diet-induced cardiac damage and fibrosis by upregulating and activating SIRT1. *Saudi J. Biol. Sci.* **2021**, *28*, 7012–7021. [CrossRef]

185. Gao, D.; Wang, H.; Xu, Y.; Zheng, D.; Zhang, Q.; Li, W. Protective effect of astaxanthin against contrast-induced acute kidney injury via SIRT1-p53 pathway in rats. *Int. Urol. Nephrol.* **2019**, *51*, 351–358. [CrossRef]

186. Zhang, D.; Zhao, L.; Wong, L. β-Nicotinamide Monucleotide Supplement with Astaxanthin and Blood Orange Enhanced NAD+ Bioavailability and Mitigated Age-Associated Physiological Decline in Zebrafish. *Curr. Dev. Nutr.* **2022**, *6*, 54. [CrossRef]

187. Petyaev, I.M. Lycopene Deficiency in Ageing and Cardiovascular Disease. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 3218605. [CrossRef]

188. Li, J.; Zhang, Y.; Zeng, X.; Cheng, Y.; Tang, L.; Hong, D.; Yang, X. Lycopene ameliorates insulin resistance and increases muscle capillary density in aging via activation of SIRT1. *J. Nutr. Biochem.* **2022**, *99*, 108862. [CrossRef]

189. Liu, X.; Dilxat, T.; Shi, Q.; Qiu, T.; Lin, J. The combination of nicotinamide mononucleotide and lycopene prevents cognitive impairment and attenuates oxidative damage in D-galactose induced aging models via Keap1-Nrf2 signaling. *Gene* **2022**, *822*, 146348. [CrossRef] [PubMed]

190. Bielak-Zmijewska, A.; Grabowska, W.; Ciolko, A.; Bojko, A.; Mosieniak, G.; Bijoch, Ł.; Sikora, E. The Role of Curcumin in the Modulation of Ageing. *Int. J. Mol. Sci.* **2019**, *20*, 1239. [CrossRef] [PubMed]

191. Zendedel, E.; E Butler, A.; Atkin, S.L.; Sahebkar, A. Impact of curcumin on sirtuins: A review. *J. Cell. Biochem.* **2018**, *119*, 10291–10300. [CrossRef] [PubMed]

192. Hu, A.; Huang, J.-J.; Li, R.-L.; Lu, Z.-Y.; Duan, J.-L.; Xu, W.-H.; Chen, X.-P.; Fan, J.-P. Curcumin as therapeutics for the treatment of head and neck squamous cell carcinoma by activating SIRT1. *Sci. Rep.* **2015**, *5*, 13429. [CrossRef]

193. Bańkowski, S.; Petr, M.; Rozpara, M.; Sadowska-Krępa, E. Effect of 6-week curcumin supplementation on aerobic capacity, antioxidant status and sirtuin 3 level in middle-aged amateur long-distance runners. *Redox Rep.* **2022**, *27*, 186–192. [CrossRef]

194. Rhoads, T.W.; Anderson, R.M. Alpha-Ketoglutarate, the Metabolite that Regulates Aging in Mice. *Cell Metab.* **2020**, *32*, 323–325. [CrossRef]

195. Chin, R.M.; Fu, X.; Pai, M.Y.; Vergnes, L.; Hwang, H.; Deng, G.; Diep, S.; Lomenick, B.; Meli, V.S.; Monsalve, G.C.; et al. The metabolite α-ketoglutarate extends lifespan by inhibiting ATP synthase and TOR. *Nature* **2014**, *510*, 397–401. [CrossRef]

196. Bayliak, M.M.; Lushchak, V.I. Pleiotropic effects of alpha-ketoglutarate as a potential anti-ageing agent. *Ageing Res. Rev.* **2021**, *66*, 101237. [CrossRef]

197. Demidenko, O.; Barardo, D.; Budovskii, V.; Finnemore, R.; Palmer, F.R.; Kennedy, B.K.; Budovskaya, Y.V. Rejuvant®, a potential life-extending compound formulation with alpha-ketoglutarate and vitamins, conferred an average 8 year reduction in biological aging, after an average of 7 months of use, in the TruAge DNA methylation test. *Aging* **2021**, *13*, 24485–24499. [CrossRef]

198. Asadi Shahmirzadi, A.; Edgar, D.; Liao, C.-Y.; Hsu, Y.-M.; Lucanic, M.; Asadi Shahmirzadi, A.; Wiley, C.D.; Gan, G.; Kim, D.E.; Kasler, H.G.; et al. Alpha-Ketoglutarate, an Endogenous Metabolite, Extends Lifespan and Compresses Morbidity in Aging Mice. *Cell Metab.* **2020**, *32*, 447–456.e6. [CrossRef]

199. Payne, A.; Nahashon, S.; Taka, E.; Adinew, G.M.; Soliman, K.F.A. Epigallocatechin-3-Gallate (EGCG): New Therapeutic Perspectives for Neuroprotection, Aging, and Neuroinflammation for the Modern Age. *Biomolecules* **2022**, *12*, 371. [CrossRef]

200. Niu, Y.; Na, L.; Feng, R.; Gong, L.; Zhao, Y.; Li, Q.; Li, Y.; Sun, C. The phytochemical, EGCG, extends lifespan by reducing liver and kidney function damage and improving age-associated inflammation and oxidative stress in healthy rats. *Aging Cell* **2013**, *12*, 1041–1049. [CrossRef]

201. Pai, P.-Y.; Chou, W.-C.; Chan, S.-H.; Wu, S.-Y.; Chen, H.-I.; Li, C.-W.; Hsieh, P.-L.; Chu, P.-M.; Chen, Y.-A.; Ou, H.-C.; et al. Epigallocatechin Gallate Reduces Homocysteine-Caused Oxidative Damages through Modulation SIRT1/AMPK Pathway in Endothelial Cells. *Am. J. Chin. Med.* **2020**, *49*, 113–129. [CrossRef]

202. Ayissi, V.B.O.; Ebrahimi, A.; Schluesenner, H. Epigenetic effects of natural polyphenols: A focus on SIRT1-mediated mechanisms. *Mol. Nutr. Food Res.* **2013**, *58*, 22–32. [CrossRef] [PubMed]

203. Jiang, S.; Huang, C.; Zheng, G.; Yi, W.; Wu, B.; Tang, J.; Liu, X.; Huang, B.; Wu, D.; Yan, T.; et al. EGCG Inhibits Proliferation and Induces Apoptosis Through Downregulation of SIRT1 in Nasopharyngeal Carcinoma Cells. *Front. Nutr.* **2022**, *9*, 851972. [CrossRef] [PubMed]

204. Ageing and Health. Available online: https://www.who.int/news-room/fact-sheets/detail/ageing-and-health (accessed on 11 November 2022).

205. Peclat, T.R.; Thompson, K.L.; Warner, G.M.; Chini, C.C.S.; Tarragó, M.G.; Mazdeh, D.Z.; Zhang, C.; Zavala-Solorio, J.; Kolumam, G.; Liang Wong, Y.; et al. CD38 inhibitor 78c increases mice lifespan and healthspan in a model of chronological aging. *Aging Cell* **2022**, *21*, e13589. [CrossRef] [PubMed]