


# Application of Native ESI-MS to Characterize Interactions between Compounds Derived from Fragment-Based Discovery Campaigns and Two Pharmaceutically Relevant Proteins

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## **Supporting Information**

### **Application of native ESI-MS to characterize interactions between compounds derived from fragment based discovery campaigns and two pharmaceutically relevant proteins**

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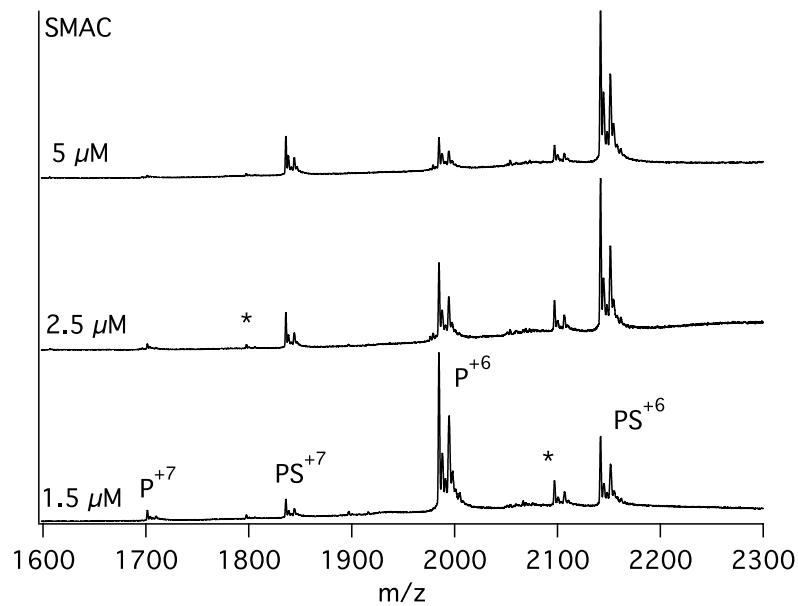
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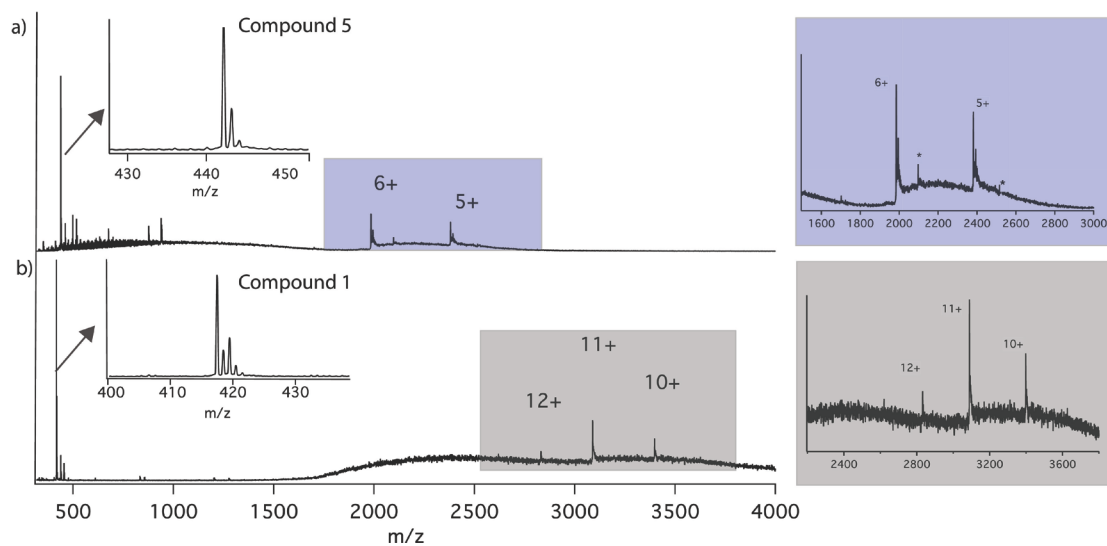
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#### **Expression and Purification of CDK2**

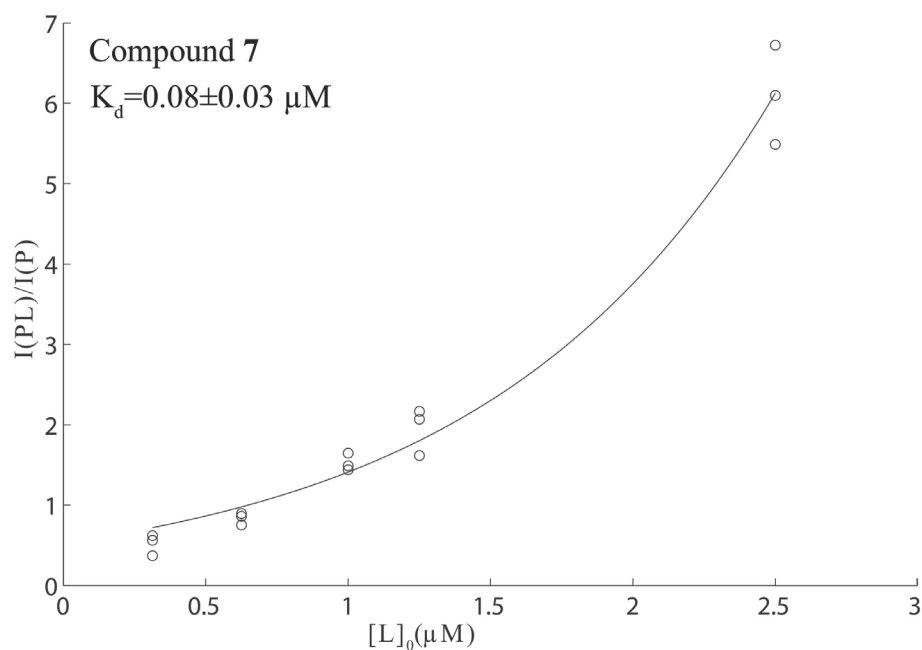
The kinase domain for human CDK2 with an N-terminal GST-tag, was transformed into E. Coli Rosetta-Gami 2(DE3) cells (Novagen). Cultures were grown for 4–6 h at 37 °C until they reached an optical density at 600 nm (OD<sub>600</sub>) of 1. At this point the temperature was decreased to 18 °C and 0.1 mM isopropyl β-D-1-thiogalactopyranoside (IPTG) was added. The cultures were allowed to grow for an additional 20–24 h at 18 °C and were harvested by centrifugation (15 min at 6000 g). Harvested cells were resuspended to a volume of 200 ml in a buffer with 50 mM HEPES, 150 mM NaCl, 2 mM dithiothreitol (DTT), pH 7.4 containing 0.5 ml protease inhibitors (Roche complete tablet) and 0.5 ml DNase and sonicated on ice for 10 minutes (5' on / 10' off). Cells were clarified by centrifugation in a JA-14 rotor at 14000 rpm for 1 hour. After sonication and centrifugation, the supernatant was purified by GST-affinity column chromatography (GE LifeSciences). After incubation of peak fractions with GST-PreScission protease (20:1) at 4 °C, the cleaved GST tag was removed by a second GST-affinity column. Fractions containing the protein were loaded onto a Superdex 75 (16/60) column, eluting with a buffer of 25 mM HEPES, 150 mM NaCl, 2 mM DTT, pH 7.4. Purified CDK2 was concentrated to ~10 mg/mL and stored at –80 °C.



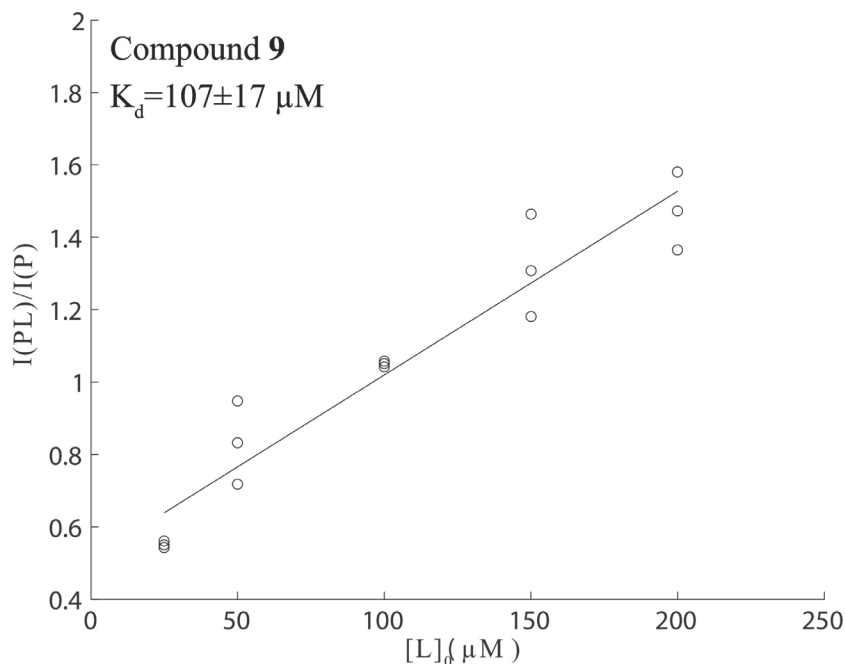
**Figure S1:** Native ESI-MS spectra of titration of SMAC peptide against 5  $\mu\text{M}$  XIAP. At increased concentration of the SMAC peptide the intensity of the peak of the XIAP-SMAC peptide complex (PS) was increased while that of the protein (P) peak was decreased. The highlighted peak (\*) in this and all subsequent XIAP spectra corresponds to the complex of XIAP and a 673 Da peptide that was a component of the bacterial growth medium. However, even at high concentrations of the SMAC peptide this peak was not depleted by competition with the SMAC peptide.



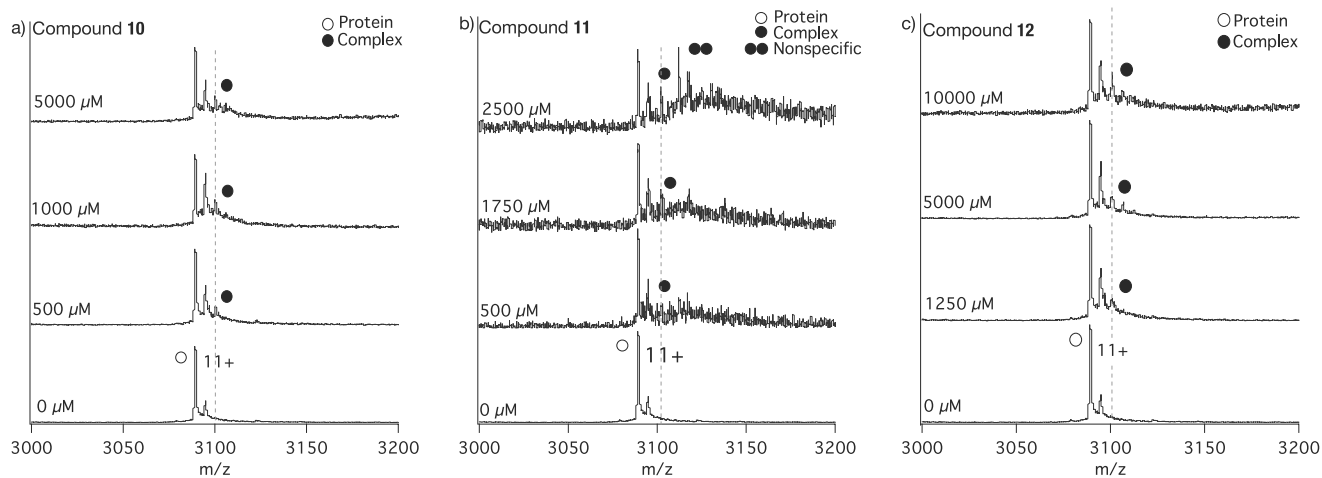
**Figure S2:** Nonspecific binding - nano-electrospray mass spectra of a) 5  $\mu\text{M}$  XIAP and 50  $\mu\text{M}$  compound **5** and b) 5  $\mu\text{M}$  CDK2 and 50  $\mu\text{M}$  compound **1**. No ligand binding to the protein was observed in either case. The most abundant peak in both spectra was that corresponding to free compound. The m/z ranges of the protein spectra are highlighted in the zoomed spectra on the right.



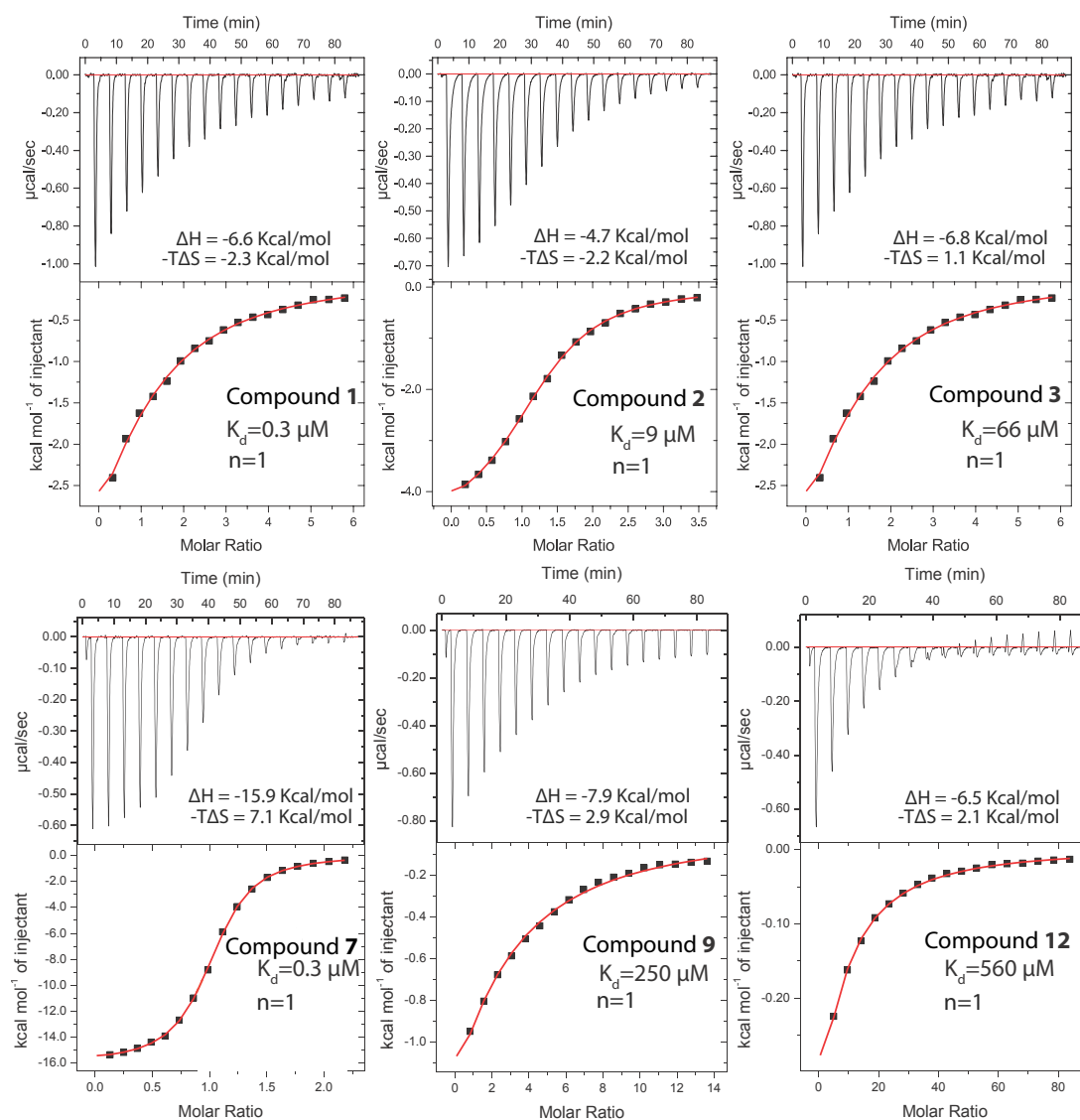
**Figure S3:** Titration curve from triplicate measurements of the compound 7 to 5  $\mu\text{M}$  CDK2 in 50 mM  $\text{NH}_4\text{Ac}$ , 1 % DMSO. The measured relative peak areas (I) of the complex (PL) to the protein (P) are plotted versus the initial concentration of the compound ( $[\text{L}]_0$ ).



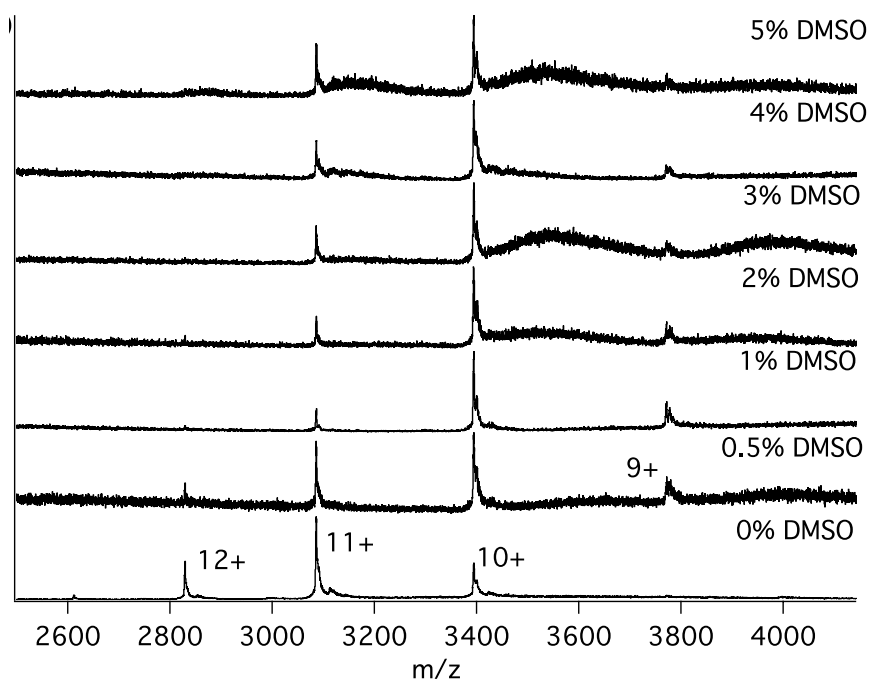
**Figure S4:** Titration curve from triplicate measurements of compound 9 against 5  $\mu\text{M}$  CDK2 in 50 mM  $\text{NH}_4\text{Ac}$ , 1 % DMSO. The measured relative peak areas (I) of the complex (PL) to the protein (P) are plotted versus the initial concentration of the compound ( $[\text{L}]_0$ ). For weak affinities with  $K_d$  values > 100  $\mu\text{M}$  the quality of the fit is reduced.



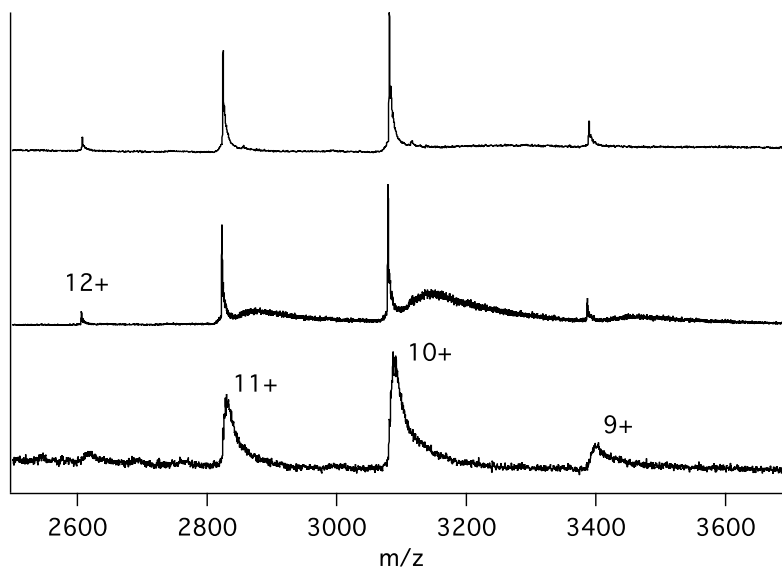
**Figure S5:** Native ESI-MS spectra for the 11+ charge state of the titration of the three compounds with the lowest affinity, **10-12** against 5  $\mu\text{M}$  CDK2 in 50 mM  $\text{NH}_4\text{Ac}$ , 1% DMSO



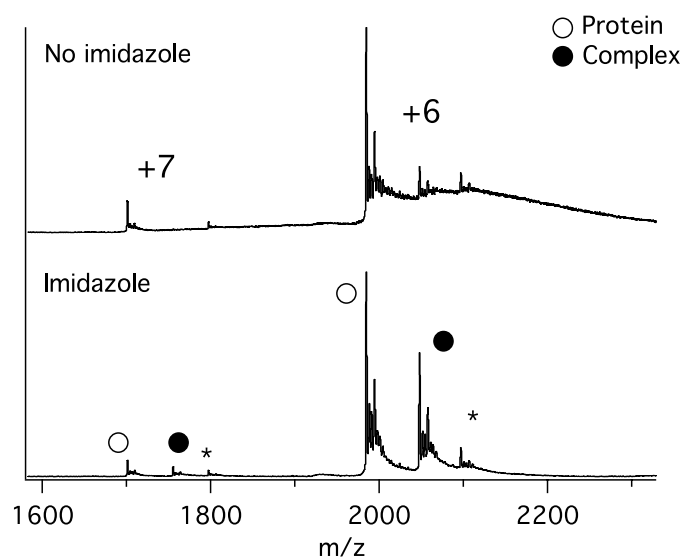
**Figure S6:** Measurement of thermodynamic parameters including dissociation constant ( $K_d$ ) and stoichiometry ( $n$ ) by ITC for interactions between XIAP with compounds **1-3** and CDK2 with compounds **7, 9, 12**.



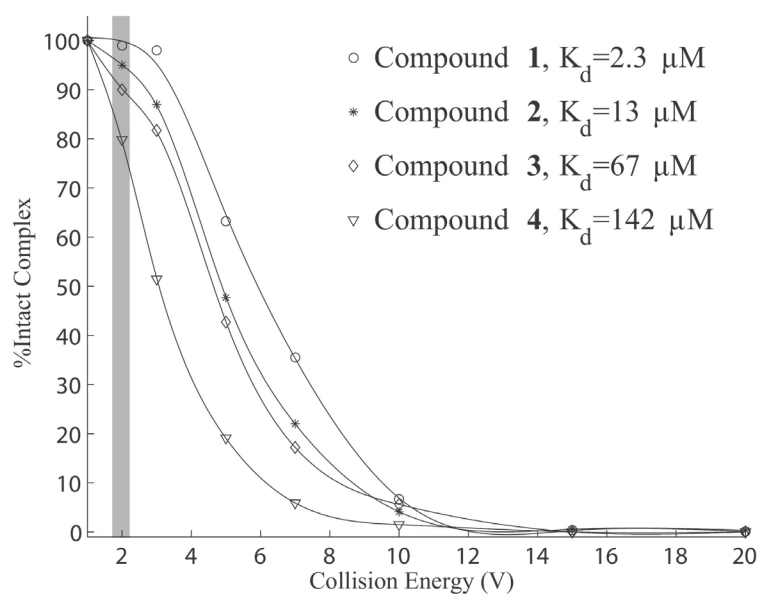
**Figure S7:** Nanoelectrospray mass spectra of 5 μM CDK2 in 50 mM NH<sub>4</sub>Ac containing 0-5 % DMSO.



**Figure S8:** Nanoelectrospray mass spectra of 5 μM CDK2 in 50 mM NH<sub>4</sub>Ac after one desalting step (lower spectrum), two desalting steps (middle spectrum) and three desalting steps (upper spectrum). The S/N ratio increased sequentially with successive desalting steps.



**Figure S9:** Representative nano ESI-MS spectra of 5  $\mu\text{M}$  XIAP in complex with 50  $\mu\text{M}$  antagonist, +/- 10 mM imidazole in 20 mM  $\text{NH}_4\text{Ac}$ , pH 7.5. The lower spectrum was recorded after addition of imidazole.



**Figure S10:** Dissociation curves for the various noncovalent XIAP complexes: relative intensity of the complex plotted as a function of the collision energy to show gas-phase stabilities of the complexes. The optimum collision energy of 2 V is highlighted.



**Table S1. Compounds, molecular weight (MW), structures, clogP, the number of heavy atoms (heavy atom count, HAC), solubility (sol) in 50 mM NH<sub>4</sub>Ac / 1% DMSO, pH 7.5 where available, and the purity (in %) of the compounds. \* Fragment hits; \*\* Compounds in the hit-to-lead phase; \*\*\* Compounds in the lead optimization phase.**

Compound	MW (Da)	Structure	Compound	MW (Da)	Structure	Compound	MW(Da)	Structure
<b>1</b> **	416	 clogP: 3.430 HAC: 29 purity 99.7 %	<b>5</b> ***	438	 clogP: 1.710 HAC: 32 sol: 770 μM purity 98.8 %	<b>9</b> *	223	 clogP: 1.090 HAC: 15 sol: 770 μM purity 89.7 % [2]
<b>2</b> **	379	 clogP: 3.876 HAC: 28 purity 95.3 %	<b>6</b> ***	360	 clogP: 2.435 HAC: 26 sol: <10 μM purity 99.6 %	<b>10</b> *	118	 clogP: 1.625 HAC: 9 sol: 4810 μM purity 98.8 %
<b>3</b> *	180	 clogP: -0.437 HAC: 13 purity 88.8 % [1]	<b>7</b> ***	381	 clogP: 0.325 HAC: 25 sol: 5100 μM purity 100 %	<b>11</b> *	144	 clogP: -0.157 HAC: 11 sol: 2890 μM purity 98.1 %
<b>4</b> **	259	 clogP: 1.751 HAC: 19 purity 98 %	<b>8</b> **	205	 clogP: 3.240 HAC: 14 sol: 190 μM purity 99.5 %	<b>12</b> *	129	 clogP: 0.435 HAC: 8 sol: 5210 μM purity 100 %

[1] Major impurity: tert-butyloxycarbonyl (BOC) protecting group

[2] Major impurity: dimer of compound 9

**Table S2:** Measured and calculated masses of the proteins and their noncovalent complexes

Proteins/Mass	Compound theoretical mass (Da)	Measured mass of noncovalent complexes (Da)	Measured $\Delta M$ = Mass of complex – Mass of protein (Da)
<b>XIAP</b> Theoretical: 11840 Da Measured: 11904 $\pm$ 2 Da <sup>1</sup>	Compound <b>1</b> : 416	12324 $\pm$ 3	420
	Compound <b>2</b> : 379	12288 $\pm$ 3	384
	Compound <b>3</b> : 180	12089 $\pm$ 5	185
	Compound <b>4</b> : 259	12164 $\pm$ 3	260
<b>CDK2</b> Theoretical: 33929 Da Measured: 33891 $\pm$ 5 Da	Compound <b>5</b> : 438	34323 $\pm$ 2	432
	Compound <b>6</b> : 360	34251 $\pm$ 1	360
	Compound <b>7</b> : 381	34271 $\pm$ 3	380
	Compound <b>8</b> : 205	34101 $\pm$ 2	210
	Compound <b>9</b> : 223	34120 $\pm$ 6	229
	Compound <b>10</b> : 118	34012 $\pm$ 7	121
	Compound <b>11</b> : 144	34035 $\pm$ 6	140
Compound <b>12</b> : 129	34023 $\pm$ 14	132	

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<sup>1</sup> The difference between the measured and theoretical mass is due to the zinc ion (65 Da).