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Draft Genome Sequence of *Klebsiella* pneumoniae 704SK6, an OXA-48- and CTX-M-15-Encoding Wastewater Isolate

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ABSTRACT The Swiss wastewater isolate *Klebsiella pneumoniae* 704SK6, encoding OXA-48 and CTX-M-15 β -lactamases, was fully sequenced. The assembly resulted in an open chromosome of 5,208,104 bp in size (G+C content, 57.6%) and four closed plasmid sequences of 209,651, 197,670, 65,998, and 63,605 bp in size.

Klebsiella pneumoniae is pervasive in the environment and an intestinal commensal. Klt is also a severe threat to human health, as it causes both nosocomial and community-acquired infections (1). OXA-48 is a widespread class D β -lactamase which is not susceptible to β -lactamase inhibitors. While it does not hydrolyze extendedspectrum cephalosporins, it degrades penicillins very efficiently and carbapenems at lower, but still clinically significant, rates (2). OXA-48 producers frequently also encode extended-spectrum β -lactamases (ESBLs), resulting in complete β -lactam resistance and treatment failure with this class of antibiotics (3).

K. pneumoniae isolate 704SK6 was isolated from wastewater near Basel, Switzerland, in December 2015 (4). The genome was sequenced at the Functional Genomics Center Zurich (FGCZ) using Pacific Biosciences (PacBio) single-molecule real-time (SMRT) technology RS2 reads (C4/P6 chemistry). *De novo* assembly was carried out using SMRT Analysis 2.3 with the HGAP3 protocol, and sequences were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (5). Sequence type (ST), acquired antibiotic resistances, and plasmid incompatibility (Inc) groups were assessed using the MLST-1.8 server (6), ResFinder 2.1 (7), and PlasmidFinder 1.3 (8), respectively (see http://www.genomicepidemiology.org/).

The chromosome of *K. pneumoniae* 704SK6 (ST437) is not fully closed. The linear sequence is 5,208,104 bp in length, with a G+C content of 57.6%, and it contains β -lactam (*bla*_{SHV-11}), fosfomycin (*fosA*), and quinolone (*oqxA* and *oqxB*) resistance genes. Assembly revealed four complete (closed) plasmid sequences, which are provided here in descending order of size: (i) p704SK6_1 (209,651 bp; G+C content, 45.2%) carries no acquired resistance genes but otherwise shows high similarity to the *bla*_{VIM}-carrying *K. pneumoniae* plasmid pKP04VIM (GenBank accession no. KU318421, 91% query coverage, 99% identity); (ii) p704SK6_2 (197,670 bp; G+C content, 52.9%; IncFII) contains aminoglycoside [*aph*(3')-*la*], macrolide [*mph*(A)], and sulfonamide (*sul1*) resistance genes and is very similar to *K. pneumoniae* plasmid p34618 (accession no. CP010393, 98% query coverage, 99% identity); (iii) p704SK6_3 (65,998 bp; G+C content, 51.0%; IncFIB) features a class 1 integron (In1407 [9]) and is a multidrug resistance (MDR) plasmid carrying aminoglycoside [*aac*(3)-*lInd*], β -lactam (*bla*_{OXA-1}, *bla*_{TEM-1B}, and *bla*_{CTX-M-15}), fluoroquinolone/aminoglycoside [*aac*(6')-*lb-cr*], tetracycline (*tetD*), and trim-

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ethoprim (*dfrA30*) resistance genes, and it most closely resembles *K. pneumoniae* plasmid p_IncFIB_DHQP1002001 (accession no. CP016810, 67% query coverage, 99% identity); (iv) p704SK6_4 (63,605 bp; G+C content, 51.2%; IncL) encodes the OXA-48 carbapenemase and is very similar to pOXA-48 (accession no. JN626286, 95.1% identical on nucleotide level). There are some notable differences between these two plasmids; the gene encoding OXA-48 lies between two IS*1999* transposases and is 100% identical in the two plasmids, but it is located on opposite strands. Also, our newly sequenced plasmid features a 504-bp insertion element, IS*1* protein InsB (accession no. WP_001119291), directly upstream (complementary strand) of its OXA-48 gene, as well as downstream (complementary strand) of its *korC* gene as part of a 776-bp insertion. Excluding these regions, the two plasmids are 99.4% identical.

Isolates like *K. pneumoniae* 704SK6 highlight the wide dissemination of pan- β -lactam-resistant strains of this species and the only slightly modified, ever-reoccurring, pOXA-48 IncL plasmid.

Accession number(s). Sequence and annotation data of the genome have been deposited in GenBank under accession numbers CP022143 (chromosome), CP022144 (p704SK6_1), CP022145 (p704SK6_2), CP022146 (p704SK6_3), and CP022147 (p704SK6_4). This is the first version of this genome.

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REFERENCES

- Gomez-Simmonds A, Uhlemann AC. 2017. Clinical implications of genomic adaptation and evolution of carbapenem-resistant *Klebsiella pneumoniae*. J Infect Dis 215:S18–SS27. https://doi.org/10.1093/infdis/jiw378.
- Poirel L, Potron A, Nordmann P. 2012. OXA-48-like carbapenemases: the phantom menace. J Antimicrob Chemother 67:1597–1606. https://doi .org/10.1093/jac/dks121.
- Poirel L, Bonnin RA, Nordmann P. 2012. Genetic features of the widespread plasmid coding for the carbapenemase OXA-48. Antimicrob Agents Chemother 56:559–562. https://doi.org/10.1128/AAC.05289-11.
- Zurfluh K, Bagutti C, Brodmann P, Alt M, Schulze J, Fanning S, Stephan R, Nüesch-Inderbinen M. 2017. Wastewater is a reservoir for clinically relevant carbapenemase and 16S rRNA methylase producing *Enterobacteriaceae*. Int J Antimicrob Agents. https://doi.org/10.1016/j.ijantimicag.2017.04.017.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.

- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.06094-11.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi.org/ 10.1093/jac/dks261.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/ AAC.02412-14.
- Moura A, Soares M, Pereira C, Leitão N, Henriques I, Correia A. 2009. INTEGRALL: a database and search engine for integrons, integrases and gene cassettes. Bioinformatics 25:1096–1098. https://doi.org/10.1093/ bioinformatics/btp105.